

Current issues and potential solutions for the electrospinning of major polysaccharides and proteins: A review

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Abstract

Biopolymers, especially polysaccharides and proteins, are the promising green replacement for petroleum based polymers. Due to their innate properties, they are effectively used in biomedical applications, especially tissue engineering, wound healing, and drug delivery. The fibrous morphology of biopolymers is essentially required for the effectiveness in these biomedical applications. Electrospinning (ES) is the most advanced and robust method to fabricate nanofibers (NFs) and provides a complete solution to the conventional methods issues. However, the major issues regarding fabricating polysaccharides and protein nanofibers using ES include poor electrospinnability, lack of desired fundamental properties for a specific application by a single biopolymer, and insolubility among common solvents. The current review provides the main strategies for effective electrospinning of the major biopolymers. The key strategies include blending major biopolymers with suitable biopolymers and optimizing the solvent system. A systematic literature review was done to provide the optimized solvent system of the major biopolymers along with their best possible biopolymeric blend for ES. The review also highlights the fundamental issues with the commercialization of ES based biomedical products and provides future directions to improve the fabrication of biopolymeric nanofibers.

Keywords:

Biopolymers; Electrospinning; Nanofibers; Biological macromolecules

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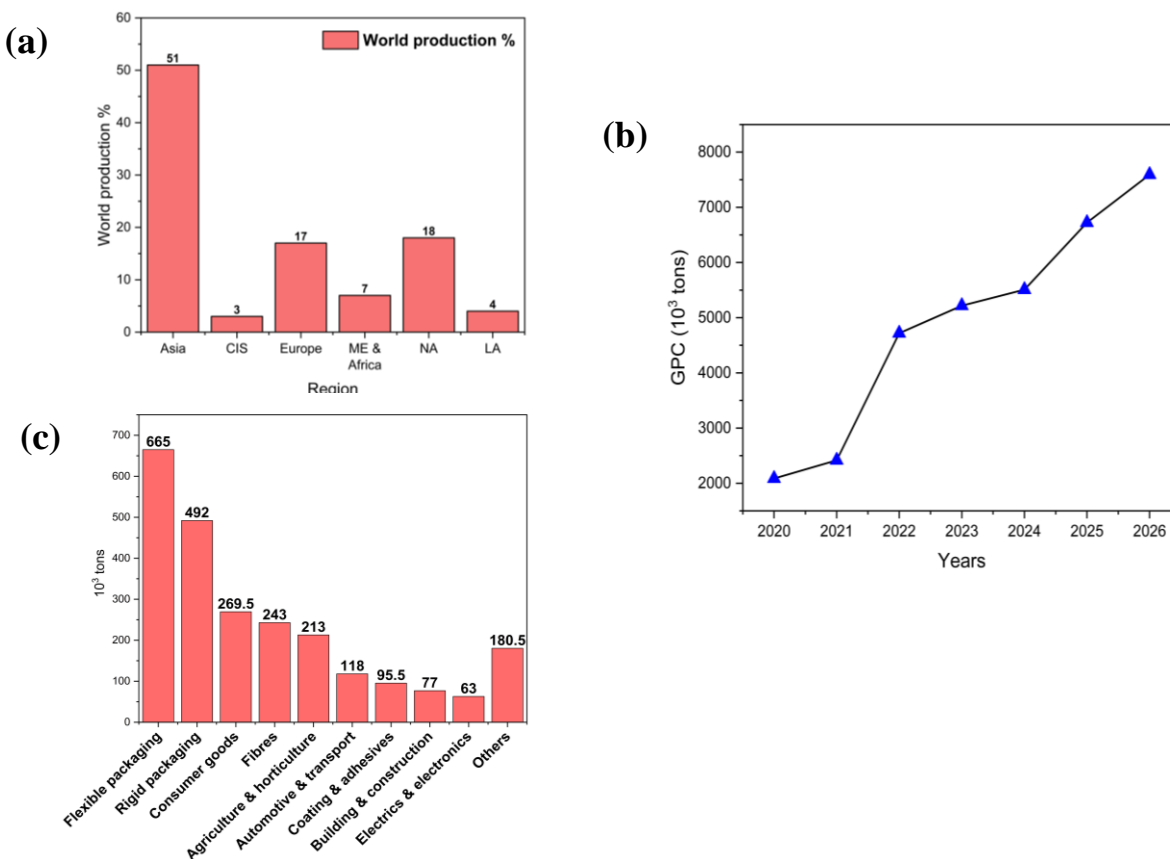
1 **1. Introduction**

2 In the mid-20th century, significant policy changes were made to discourage using petroleum-
3 based polymers. The main reasons for this initiation were the recycling and hazardous issues
4 associated with petroleum based polymers [15]. However, the industry continues to grow as it
5 employs almost 1.6 million people in Europe, and the continent's total annual revenue is roughly
6 360 billion euros [16]. There has been a progressive rise from 2 million tons of plastic
7 manufacture in 1950 to 380 million tons in 2015; just 18% of this was recycled. An estimated
8 12.5 million tons of plastic made their way into the ocean in 2010, mainly from nations around
9 the coast [19]. Fig. 1a shows the percentage of world percentage production of plastics based on
10 different regions [20].

11 Irrespective of the production and growth of the plastic industry, synthetic polymers are
12 becoming a constant threat to the environment [21]. Due to their nondegradable nature, these
13 plastic materials poison the environment when disposed of in landfills or incinerators, whereas
14 tertiary recycling methods include chemical, thermochemical, and pyrolysis [22]. However,
15 reusing all petroleum based plastics is difficult because of their environmental toxicity and
16 recycling difficulties [23]. Above all, these petroleum based polymers are not suitable at all for
17 any biomedical application since they are toxic and non-biodegradable. All these issues caused
18 the researchers to look for alternatives [24]. Researchers increasingly focus on environmentally
19 benign, biodegradable, and readily accessible materials instead of petroleum based polymers.
20 The most promising alternatives to these petroleum based polymers are biopolymers [25]. In the
21 past decade, a sharp increase in industrial and research fields has been observed for biopolymers
22 (Fig. 1c). Biopolymers are the polymers produced by living things and can be degraded
23 biologically. Biopolymers are taking over the market from synthetic polymeric materials. These

24 compounds, such as biopolymers, bioplastics, vitamins, and organic acids, may be made from
 25 natural resources using biotechnology [26].

26 About 1.1% of plastic output worldwide comes from bioplastics, with the European Union (EU)
 27 leading the charge [32]. Biopolymers have seen a steady rise in investment over the last several
 28 years. A recent survey shows that 1% of the world's plastics are produced from biopolymers
 29 [27]. The report also estimated that biopolymers' global production capacity (GPC) would be
 30 increased by 214.15% by 2026 (Determined by the authors based on data [https://ect-](https://ect-center.com/blog/biodegradable-polymers)
 31 [center.com/blog/biodegradable-polymers](https://ect-center.com/blog/biodegradable-polymers)) (Fig. 1b).



*CIS: Commonwealth independent states; ME: Middle east; NA: North America; LA: Latin America

Fig. 1 Production comparison of polymers, (a) Plastic production in major regions, (b) Prediction of biopolymers production, (c) Utilization of biopolymers in different industrial sectors

32 Biopolymers can be divided into three distinct classes depending on the origin of their raw
33 materials. The three classes include (1) natural biopolymers, (2) synthetic biopolymers, and (3)
34 microorganism-based biopolymers (Fig. 2) [28]. Some of the most diverse polymeric
35 macromolecules with exceptional structural and metabolic properties may be found in
36 microorganisms, including bacteria, fungi, yeasts, molds, smuts, and many more forms of what
37 we consider "primitive life" [29]. These biopolymers comprise various polysaccharides and
38 proteins, including cellulose, chitin, chitosan, dextran, hyaluronic acid, silk, keratin, and many
39 more. For a long time, plant-based polysaccharides and proteins have been valuable and
40 sustainable resources [30]. All living organisms ranging from microorganisms to highly evolved
41 terrestrial, aquatic, or aerial animals, may produce natural structures with great potential in
42 biomedicine [31]. These structures are present in a large variety, from glycosaminoglycans (such
43 as chitin and hyaluronic acid) and proteoglycans to proteins (majorly collagen, elastin, gelatin) to
44 the genetic material deoxyribonucleic acid (DNA) [32].

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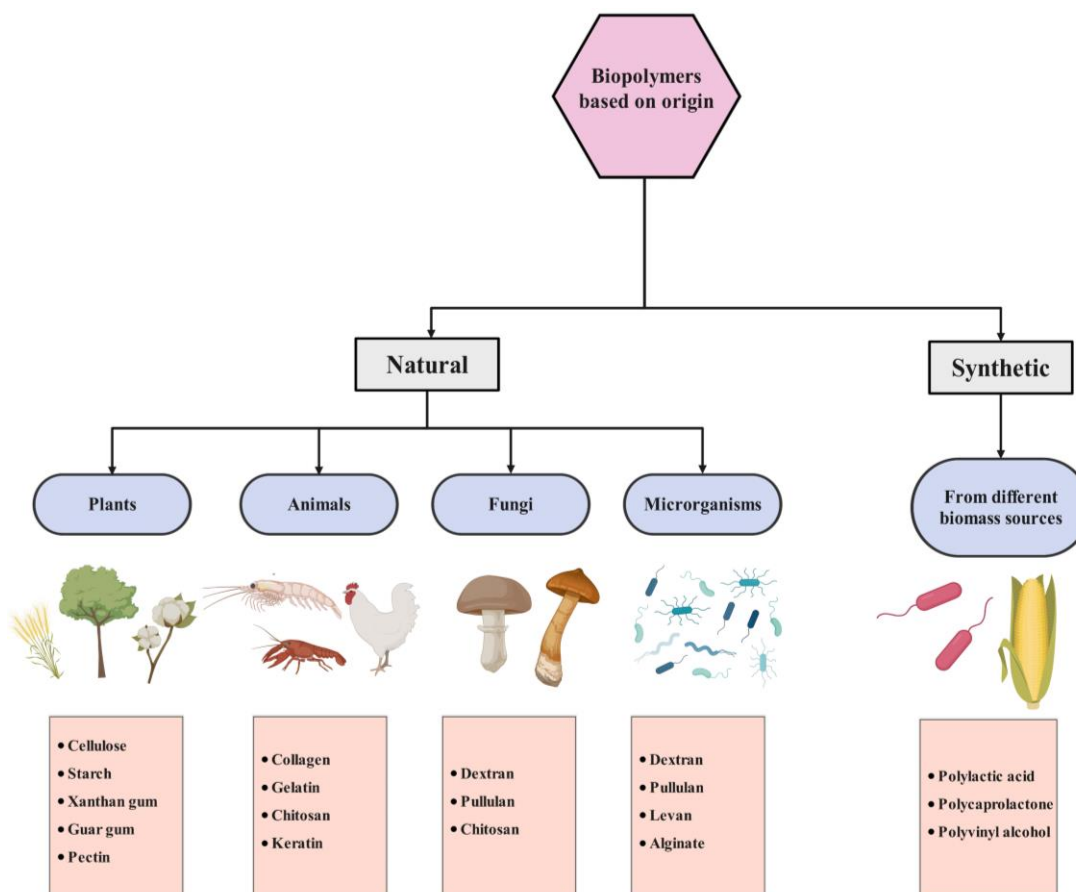


Fig. 2 Types of biopolymers based on the origin

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47 This review focuses on the electrospinning of major biopolymers and highlights the issues and
 48 the potential solution of each biopolymer. The review highlights three main strategies to
 49 overcome biopolymers ES, including chemical modifications, blending with other biopolymer,
 50 and optimizing the solvent system for the major biopolymeric combinations. The review also
 51 highlights the ES insights of the two unexplored polysaccharides, levan, and kefiran, which
 52 could be useful in future biomedical applications.

53 **1.2 Biopolymers in biomedical applications**

54 Biopolymers occur naturally in the human body and play vital roles, from enclosing cells to cell
 55 communication. They regulate the skin's moisture and elasticity to keep it in its native state.

56 They lubricate body joints and the digestive system and prevent infection by building up mucus
57 linings in the eyes and airways of humans [33]. All the significant events at the cellular level of
58 mammals revolve around the extracellular matrix (ECM), as it contains collagen and elastin. It
59 provides a high surface area based habitat for cells and tissues by providing cell adherence,
60 controlling cell signaling, and promoting cell differentiation [34]. Hence, mimicking the natural
61 ECM is the fundamental requirement of biomedical applications [35].

62 The most challenging part of fabricating a scaffold for tissue engineering is ECM replication and
63 creating a tissue-specific scaffold that meets the requirements of natural biocompatibility,
64 biodegradability, and high mechanical strength [36]. The other additional requirements of the
65 biopolymeric scaffolds include signal transportation for maintaining cell growth, skin hydration,
66 and pliability maintenance, a barrier for pathogens protection, less friction via lubrication, and so
67 on [37, 38]. However, the conventional techniques lack the fundamental mimicking of the ECM
68 [39].

69 The essential prerequisite for any drug delivery system is the controlled release of a therapeutic
70 agent. In addition, the drug carrier should prevent the therapeutic substance from being degraded
71 by enzymes before it is released slowly [40]. The drug carrier must be nontoxic, biodegradable,
72 and, most critically, should be released from the body without any adverse effect after effective
73 delivery. However, most biopolymers lack natural interaction, and the system lacks mechanical
74 strength to survive in the physiological environment [41]. Conventional drug delivery systems
75 usually use a crosslinker to overcome this issue, but the crosslinkers are mostly toxic, limiting
76 the options for developing a drug delivery system [42].

77 Biopolymers also play a significant role in wound healing by replacing traditional dressings.
78 Since traditional wound dressings were made from simple coverings, such as plant fibers or
79 animal fats, and were used to prevent further bleeding [37]. Modern technological advances have

80 allowed the fabrication of artificial materials for wound dressings with a wide range of
81 applications. Quick hemostasis and antimicrobial efficacy are two must-haves for every
82 contemporary wound dressing. The primary role of wound dressing is to stop bleeding, which
83 must be antibacterial to prevent infections from spreading [43, 44]. However, for treating major
84 wounds, a sustained drug release and high surface area to volume ratio to promote growth and
85 differentiation of the cells is the major requirement. Conventional wound healing dressings
86 mostly lack these properties owing to the methods for fabrication [45].

87 **1.3 Electrospinning**

88 The major issues highlighted in all three biomedical applications can be resolved using an
89 advanced, more flexible method to fabricate fibrous morphology [46]. Since the fibrous
90 morphology provides cellular-control properties, high porosity, better mechanical performance,
91 and high surface to mass and volume ratios compared to alternative morphologies [47]. Scaffold-
92 cell interactions may occur over a more extensive region due to the high surface to area and
93 volume ratio. The manufacturing procedure determines the fiber diameter, which may range
94 from nanometers to millimeters, allowing for the regeneration of multi-layered tissues [48]. The
95 potential biological uses of fibrous scaffolds are broadened by their malleable nature, the
96 simplicity with which their structures can be altered, and the precision with which they may be
97 shaped [49].

98 There are many techniques for the fabrication of nanofibers. However, the most promising and
99 flexible technique to fabricate nanofiber is electrospinning (ES). ES uses the blend of
100 biopolymers in a solvent system and fabricates nanofibers with the help of voltage. It provides
101 the flexibility of using a unique combination of biopolymers without any natural interactions,
102 which was challenging to achieve with old methods [50]. Incorporating unique combinations of

103 biopolymers provides the nanofibers with many critical properties crucial for biomedical
104 applications [51]. Many ES techniques are currently being used to fabricate different nanofibrous
105 morphologies for various biomedical applications. Some major ES techniques are listed in Table
106 1, along with their recent applications, and Fig. 3 shows the various morphologies of
107 biopolymers that can be achieved by using electrospinning and can be utilized for a particular
108 application.

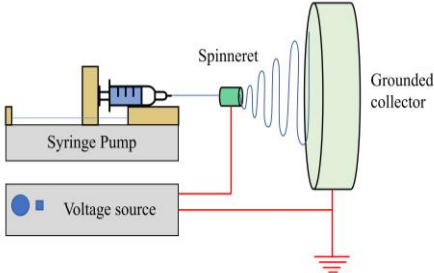
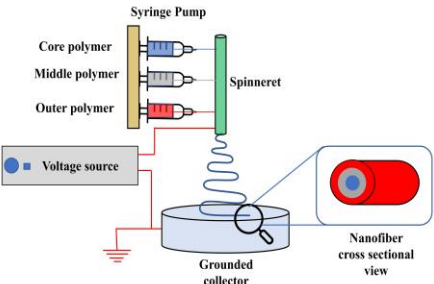
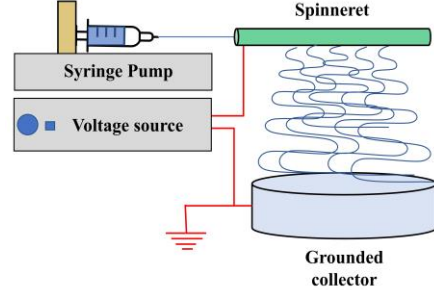
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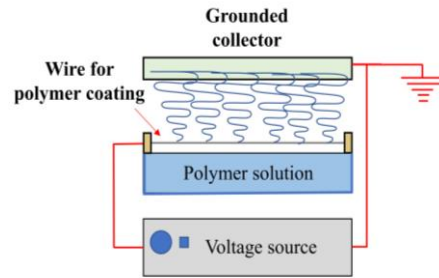
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113 Table. 1 Major strength and drawbacks of various electrospinning techniques

Electrospinning methods	Scheme of setups	Strengths	Drawbacks	Recent applications	References
Single needle (emulsion/blend)		High and uniform drug distribution, a high initial burst	The bioactivity of therapeutics is lost due to due voltage and solvent; an initial high dosage burst can cause toxicity	1. Chitosan/ polyhydroxy butyrate based nanofibers for tissue engineering 2. Alginate/ chitosan and polycaprolactone based nanofibers for capsaicin drug delivery for cancer treatment	[52, 53]
Co/tri axial		Hollow fibers formation, two separate solutions for the outer and inner core, saves the therapeutic from the solvents to dissolve biopolymers; the final structure is optimal for sustained drug delivery since the encapsulation of soluble drugs in core	Consideration of variables like interfacial tension and viscoelasticity makes the procedure designing complex	Polycaprolactone/chitosan/poly vinyl alcohol shell with resveratrol loaded core for tissue engineering	[54]
Multineedle		Enhanced rate of production	Non-uniform fiber production, electrostatic interaction of needles which are side by side	No recent potential biomedical application was observed	[55]

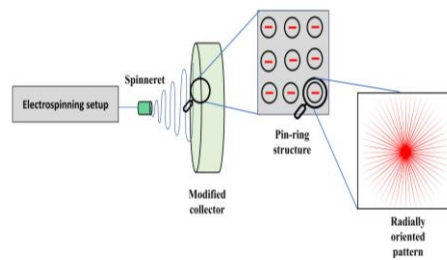
Needleless

Superior production rate as compared to needle based techniques, no spinneret clogging issue, viscous natured biopolymers can be used

High voltage, non-uniform diameter distribution of fibers, high cost, and random generation of Taylor cones

Gelatin/ polycaprolactone based nanofibers for ibuprofen sustained release

[56]

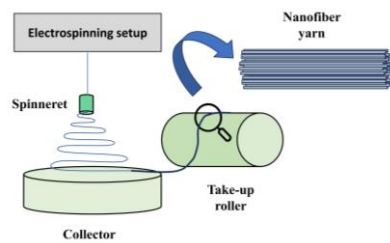
Radial nanofibers

Fabricated nanofibers have specific patterns providing biological cues for cells growth and differentiation

The process used a ring and pin structure in the collector while the area without the structure still has random nanofibers, so overall, the technique is complex with low throughput

Poly(lactic acid)/gelatin based radially aligned nanofibers for chronic wound healing

[57]

Yarn based nanofibers

The resultant nanofibers show exceptional mechanical strength, surface area and pores mimicking natural extracellular matrix for cell culture

The process is complex as it requires specialized and complex collectors and is not suitable for all biopolymers, limiting the options

Polycaprolactone based nanofiber yarns for tendon repair

[56]

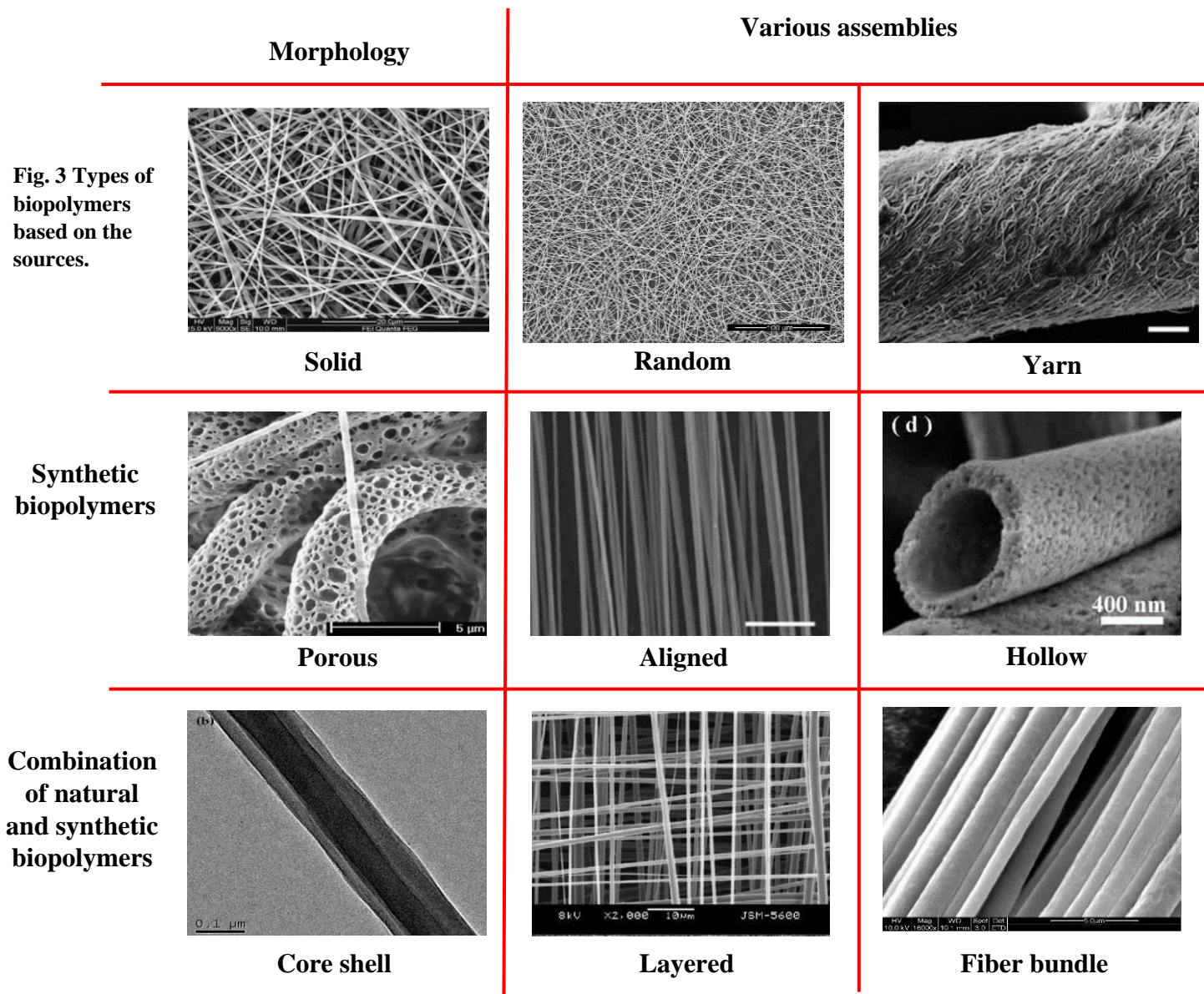


Fig. 3 Various morphologies and assemblies of natural, synthetic or combination of biopolymers [6]

115 **1.4 Importance of biopolymeric nanofibers**

116 For the application of tissue engineering, in addition to the previously mentioned features,
 117 biopolymeric nanofibrous scaffold also enhances cell adherence by including specific patterns,
 118 such as RGD (arginine, glycine, and aspartate tripeptide) [58]. All of the listed features
 119 encourage the growth and proliferation of tissues. The manufacturing process fabricating specific

120 fibers varying in size from nanosized to millimetric may be employed to recreate hierarchical
 121 tissues at various levels. Fig.4a shows the schematic representation of how the cells interact with
 122 various other factors and therapeutic agents in a nanofibers based scaffold. Fig. 4b shows the
 123 SEM micrographs of the nanofibrous yarns developed by Cai et al., (2012) [7]. The study
 124 employed human embryonic stem cell-derived mesenchymal stem cells to be developed in the
 125 scaffold. The cells changed from a round to a spindle shape, and at around 6 weeks mark a 3D
 126 bony tissue was observed around and inside the scaffold. The study showed how nanofibrous
 127 scaffolds can guide the development of bone formation.

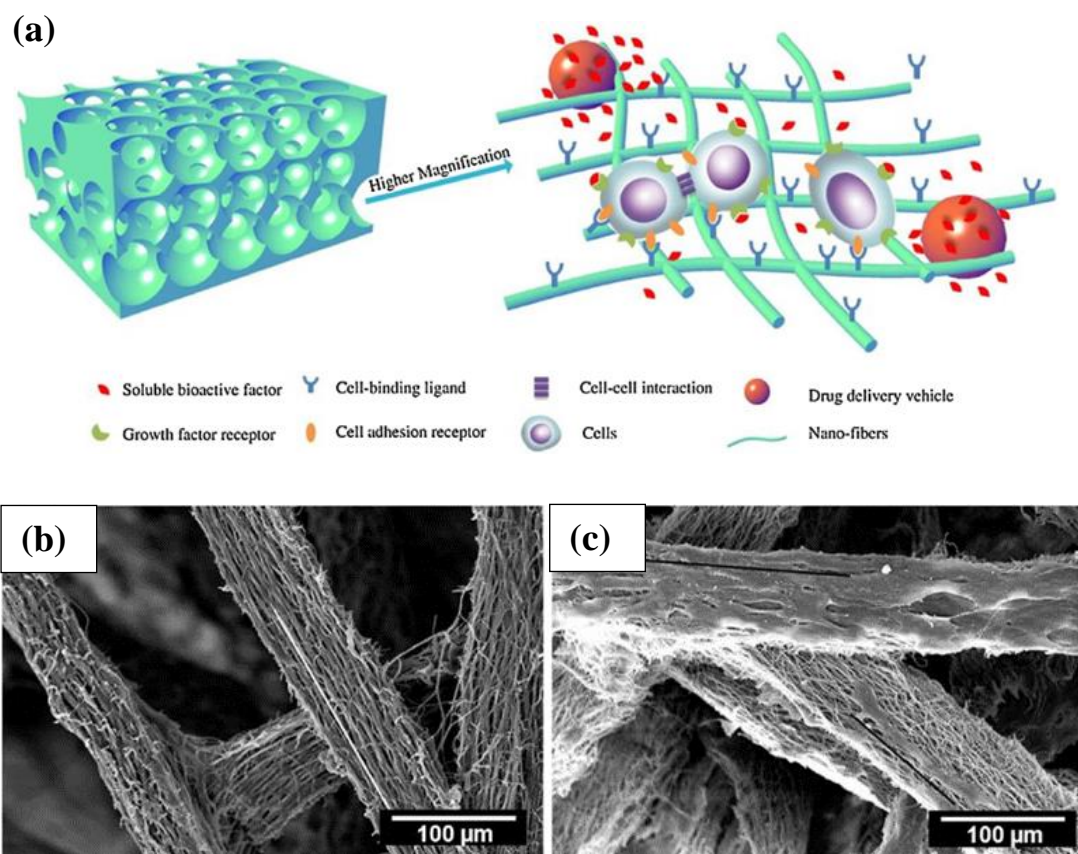


Fig. 4 Nanofibers based scaffolds for tissue engineering, (a) Schematic illustration, (b-c) SEM images at various magnifications [7]

129 For the application of drug delivery, nanofibers provide a potential solution for sustained drug
130 delivery and overcoming the issue of burst release. The achievement is only possible due to the
131 fibrous morphology, which increases the surface area to volume ratio [59]. The other more
132 effective approach is encapsulating therapeutic to provide protection and sustained release.
133 Coaxial electrospinning produces nanofibers with core-shell morphology and encapsulates drugs
134 and other therapeutics in the core to form smart drug delivery systems [60]. Abdolbaghian et al.,
135 (2022) [2] developed PLA/PVA/PVP based core-shell nanofibers for sage drug delivery. The
136 core consisted of the PLA and sage, while the shell comprised PVP and PVA (Fig. 5a & b). They
137 compared drug release of the core, shell, and core-shell structures. Their results showed that the
138 drug release was reduced by 35% in the core-shell structure compared to the dispersed drug
139 loaded structures (Fig. 5c).

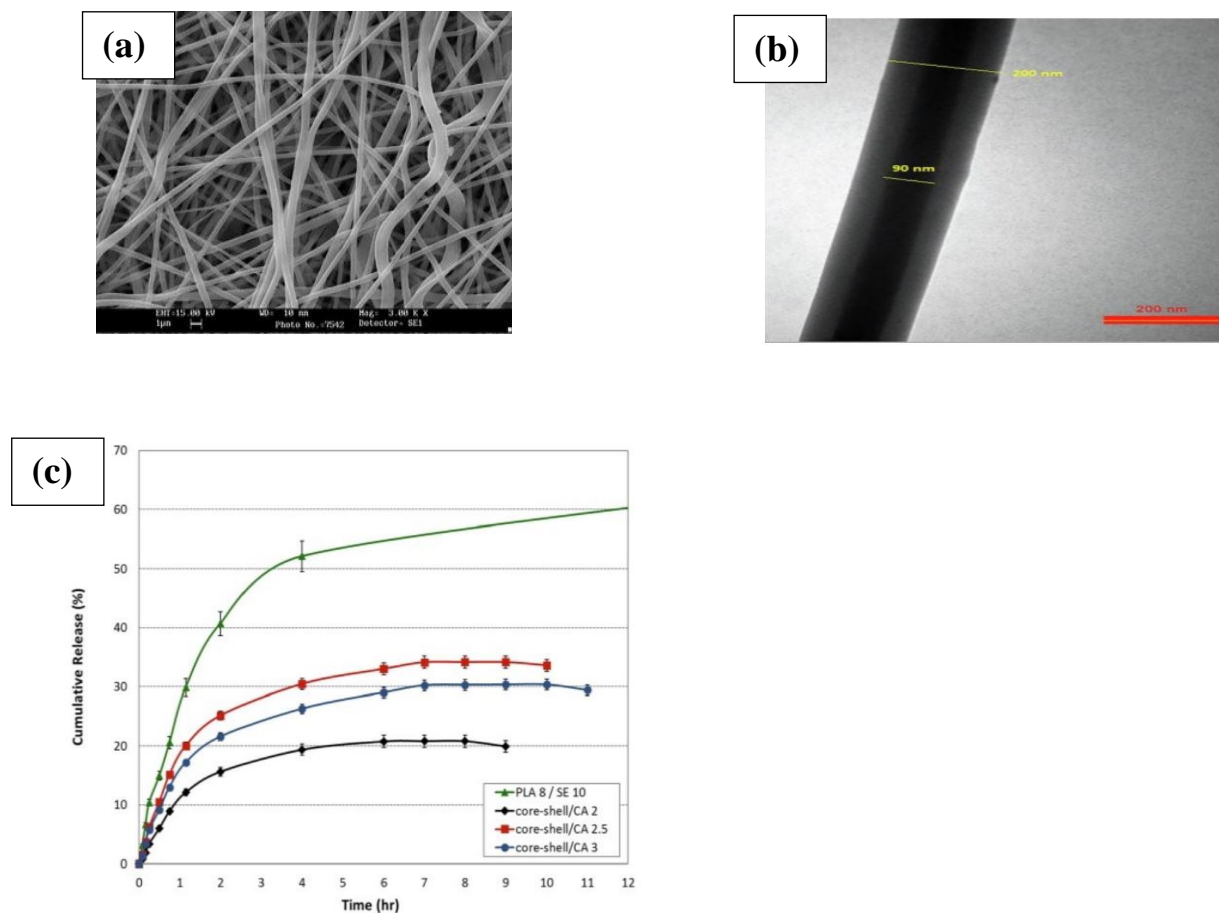


Fig. 5 Core-shell nanofibers based drug delivery system (a) Overall biocomposite SEM image, (b) Single nanofiber TEM image, (c) Drug release comparison between core-shell and simple morphology [2]

140 Nanofibers can also play a vital role in fabricating smart wound dressing since the ES allows the
 141 combination of any biopolymers, including the therapeutic, which helps advance wound healing
 142 [45]. The traditional dressing comprises polymers with no antimicrobial activities, but the
 143 biopolymeric nanofibers based dressing is innately antimicrobial. The other important concern
 144 about chronic wound healing is that the dressing can not be replaced daily due to the danger of
 145 wound infection and exhaustion of the therapeutic. This change of dressing results in disturbing
 146 the wound healing process. The nanofibrous based smart wound healing system reduces the
 147 frequency of dressing changes since there are fewer chances of wound infection, and with
 148 sustained drug release, the drug is not finished too early [61]. Due to the wide variety of options
 149 for fibrous scaffolds, the ability to adjust fiber orientation in the scaffold, and the flexible

150 structural modification, the field of biomedical applications has been greatly widened [62]. In a
151 study by Wu et al. (2022) [57], a wound healing patch for diabetic patients was developed. The
152 developed patches utilized the effect of patterns on the differentiation and growth of the cells.
153 They fabricated nanofiber and hydrogel-based bi-layered dressing consisting of the radially
154 aligned nanofibers of gelatin and polylactic acid in a radial manner (Fig. 6). The developed
155 patches showed better cell recruitment and guidance ability for better differentiation of the
156 human dermal fibroblasts (HDFS). Fig. 6a & b shows the graphical representation and
157 fluorescent imaging of the migration of HDFS along the radially aligned nanofibers from day 3
158 to 7. Fig. 6c shows the cell viability test for the developed patches and confirms the successful
159 differentiation environment mimicking the natural ECM.

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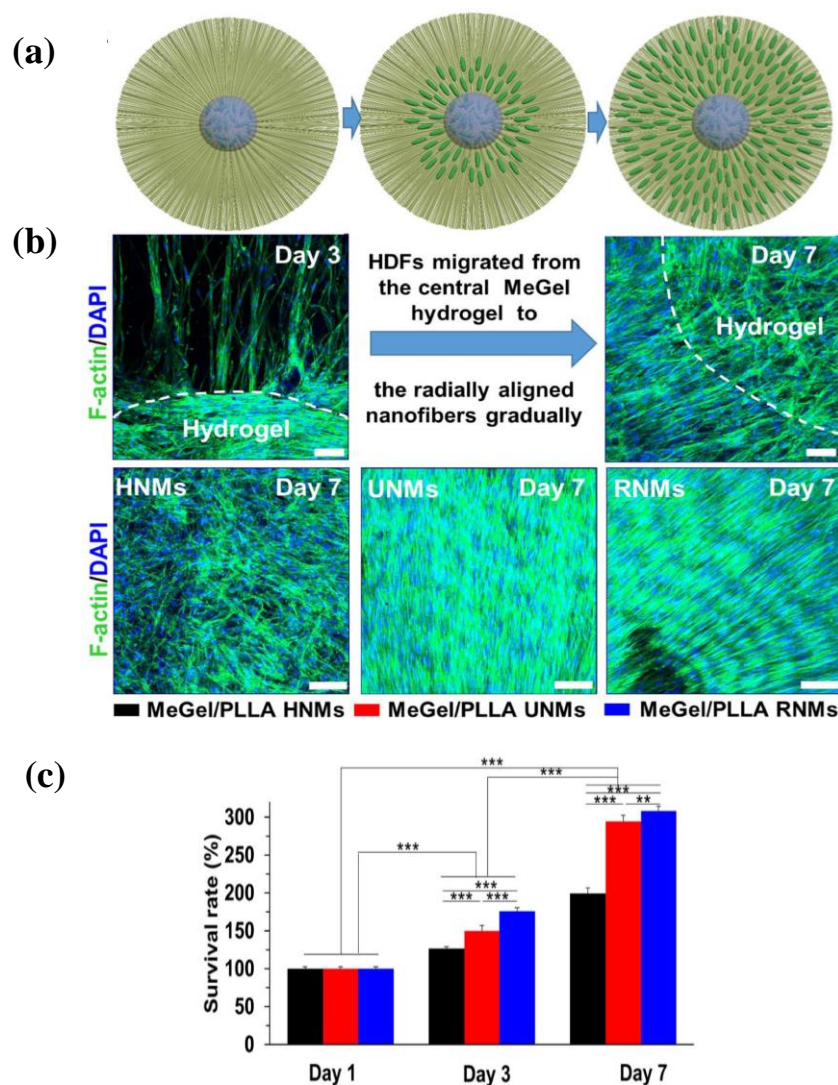


Fig. 6 Radially aligned nanofibers based wound healing patches, (a) Graphical representation, (b) Fluorescent microscope analysis, (c) Cell viability test [4]

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171 2. Major biopolymers for nanofibers fabrication

172 The major classes of biopolymers for major biomedical applications include polysaccharides,
 173 proteins, and phospholipids due to their abundance, cost-effectiveness, and high level
 174 biocompatibility [63]. However, using only a single natural biopolymer has various drawbacks
 175 and difficulties in forming a system for a biomedical application. The fundamental issue is that a
 176 single biopolymeric system cannot manifest desired properties for the nanofibers to accomplish

177 the objective of biomedical applications as they lack the whole spectrum of innate properties
178 required by a particular system including biocompatibility, cell adherence, mechanical strength
179 and antimicrobial effect [64]. The other major biopolymer issues are their insolubility and innate
180 low electrospinnability potential. There are two main approaches to overcoming the
181 electrospinnability and properties issue: biopolymers are chemically modified or blended with
182 other biopolymers for ES [65]. The blending provides the resultant nanofibers with additional
183 features for biomedical applications. Some of the major biopolymers used for ES, along with
184 their merits and demerits, are listed in Table. 2

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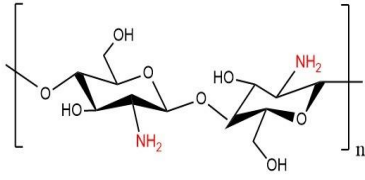
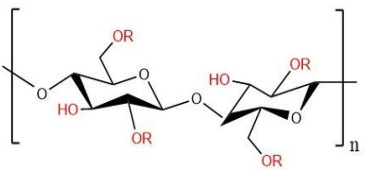
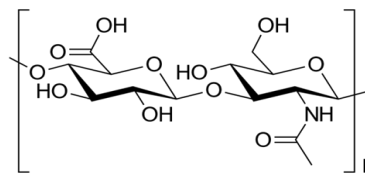
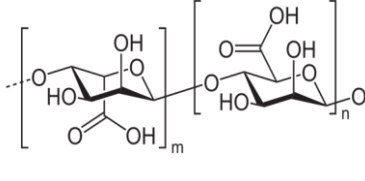
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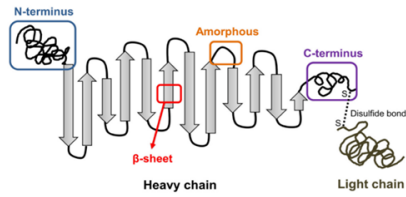
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191 Table 2. Merits and demerits of commonly used biopolymers in electrospinning

Type	Biopolymers	Chemical structures	Advantages	Disadvantages	References
Natural	Chitosan		Incorporates antimicrobial & antioxidation activity, regulates inflammation, promotes hemostasis	Less solubility and biodegradation rate are uncontrollable	[66-68]
	Cellulose		Improves thermal stability and scaffolding for the therapeutics	Highly immunogenic, less solubility and limited water holding capacity	[69-71]
	Hyaluronic acid		Natural component of the body and actively involves in the clot formation, inflammation, proliferation, and re-epithelialization	Highly viscous even at low concentration	[72-74]
	Alginate		Structure similarity with extracellular matrix, abundant availability and cost effective	High viscosity and immunogenic response	[75]

Silk fibroin

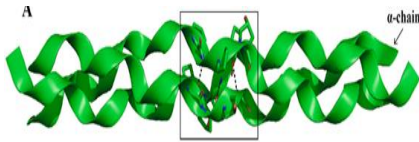


Enhanced mechanical properties, rate of biodegradation is controllable, improve water, and oxygen permeability

Extraction process is tedious, and degumming affects mechanical properties

[76]

Collagen

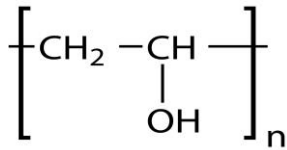


Less immunogenic, better biocompatibility, good for cell proliferation and adhesion

Sudden breakdown during degradability and extension

[77, 78]

Polyvinyl alcohol



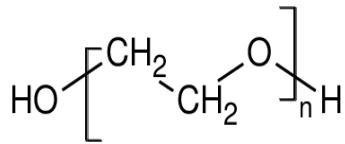
Bio-adhesive, nontoxic and better chemical resistance

Bio-inertness and low mechanical strength

[79, 80]

Synthetic

Polyethylene oxide

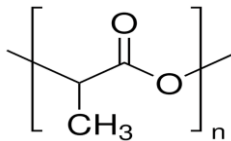


Good viscoelastic behavior in physiological environment, improves the electrospinnability as easy to blend with natural biopolymers

Non-biodegradable within the biological time frame

[81]

Polylactic acid



Excellent electrospinnability, good mechanical strength

Hydrophobic in nature and can cause inflammation and results in immunogenicity

[82, 83]

193 **2.1 Influences of the solvent system in nanofibers production**

194 The solubility issue of biopolymers is more complex than the other mentioned issues, as the
195 solvent system should not only makes the homogenous blend but also fabricates nanofibers.
196 Since it is evident in the next examples, sometimes the system will dissolve the biopolymers, but
197 the nanofibers will not be fabricated, and the ES will result in bead formation or spraying. Each
198 combination of biopolymers requires an optimized solvent system to fabricate nanofibers. The
199 properties of the solvent system also play a crucial role in the fabrication of nanofibers and their
200 morphology. The major properties of the resultant solvent system include the dielectric constant,
201 polarity, surface tension, and viscosity [84]. Georgiadou et al., 2014 [3] showed the effect of the
202 different solvent systems on the morphology of the same concentration of polylactic acid. Out of
203 the six different solvent systems, all were dissolving PLA, but only acetone/dimethylacetamide
204 formed the uniform nanofibers. The rest solvent systems either failed to fabricate the nanofibers
205 or formed the beads (Fig. 7)

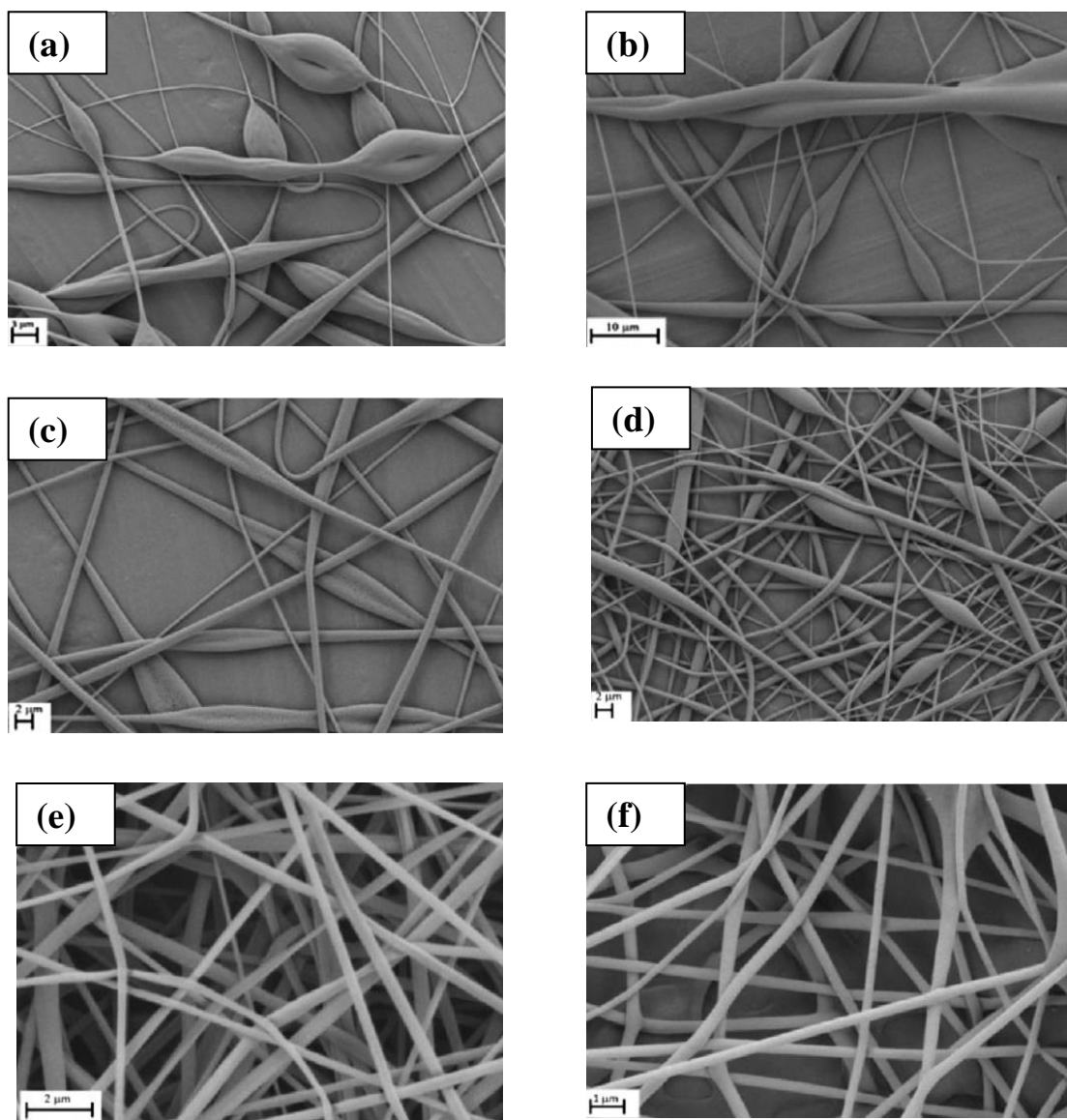


Fig. 7 Effect of different solvent systems on the nanofibers morphology of polylactic acid (10% w/v), (a) acetone/1,4-dioxane, (b) acetone/tetrahydrofuran, (c) acetone/dichloromethane, (d) acetone/chloroform, (e) acetone/dimethylformamide, (f) acetone/dimethylacetamide [3]

206 Another study by Choktaweasap et al. (2007) [6] demonstrated the effect of various solvent
 207 systems (Glacial acetic acid (GAA) (Fig. 8a), GAA/2,2,2-trifluoroethanol (TFE) (Fig. 8b),
 208 GAA/dimethyl sulfoxide (DMSO) (Fig. 8c), GAA/ethylene glycol (EG) (Fig. 8d), and
 209 AA/formamide (F) (Fig. 8e) on the morphology of gelatin NFs. They conducted the ES process
 210 with the same parameters (voltage: 7.5 kV and the distance between needle and collector 7.5 cm)

211 and the same biopolymer (gelatin) but changed the solvent system to see the direct effect of
 212 solvent systems on the final NFs. Different solvent systems resulted in the change in properties
 213 of the solution, like surface tension and conductivity, and hence the change in morphology and
 214 the diameter of the NFs was observed.

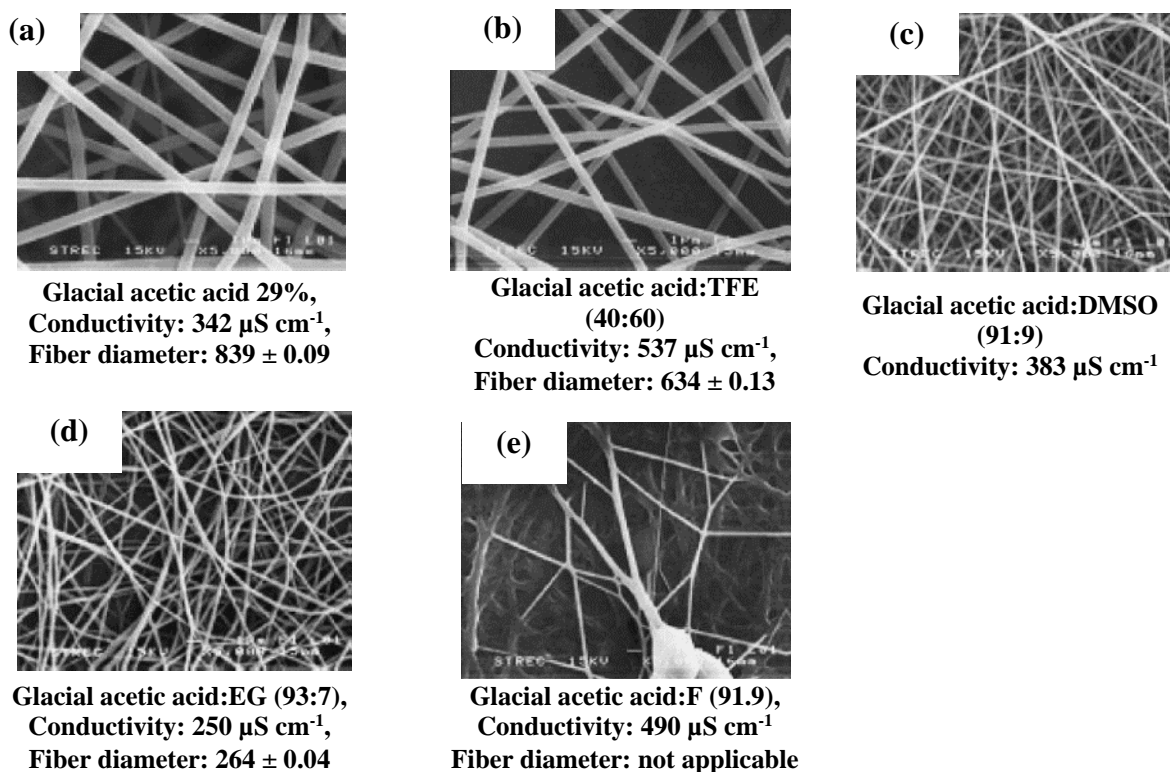


Fig. 8 Effect of different solvent systems on the gelatin nanofibers, (a) Glacial acetic acid, (b) GAA:TFE, (c) GAA:DMSO, (d) GAA:EG, (e) GAA:F [5]

215 The following section highlights the major issues with the fabrication of nanofibers for major
 216 biopolymers. It also provides potential approaches to solve those issues, which mainly include
 217 blending the biopolymer or developing an optimized solvent system. It also enlists the possible
 218 combinations of various biopolymers and the particular solvent system for a specific biomedical
 219 application.

221

222 **2.2.1 Cellulose and their derivatives**

223 According to the International Union for the Conservation of Nature (IUCN), cellulose is the
224 most abundant biopolymer, which is a homopolymer of glucose units homopolymer which is
225 connected by (1,4)-linkages [85]. Cellulose is biodegradable, bio-friendly, nontoxic, cost-
226 effective, and has high mechanical strength and chemical resistance since insoluble in most polar
227 and nonpolar solvents [86]. Cellulose source, the extraction process, polymerization (DP) degree,
228 and treatment procedure may determine cellulose polymorphism (i.e., I, II, III, and IV). DP
229 mostly depends on the source type, but the overall molecular structure stays constant and
230 significantly impacts cellulose viscosity, mechanical characteristics, and dissolving properties.
231 Thus, these three characteristics determine cellulose electrospinning and the application of its
232 fiber form [87]. These will be the advantages of employing cellulose in the nanofibers.
233 Developing technology for cellulose extraction from waste items such as food, agricultural, and
234 textile waste further increases its sustainability and promotes a green environment [88].

235 Despite the many advantages of cellulose, the major hindrances are the non-solubility of
236 cellulose. Hence, it is mainly modified into various derivatives that somehow solve the solubility
237 issue and improves the chances of electrospinnability. Most alterations are made to cellulose for
238 its better affinity for polar/nonpolar molecules [89, 90]. The advantages and disadvantages of
239 significant cellulose derivatives for ES and their common solvent systems are listed in the Table.
240 3. Fig. S1 shows the morphology of the major cellulose derivatives blending with other
241 biopolymers [8, 91, 92]

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245 **Table 3. Merits and demerits of common cellulose derivatives**

Derivative	Advantages	Solvent system	Disadvantages	Potential applications	References
Cellulose acetate	Better thermal stability, solubility, biocompatibility	Acetone: dimethyl acetamide (2:1 v/v)	Low Maximum operating temperature (40°C), Low pH working range (3-6)	Drug delivery, wound healing, microfiltration	[93-95]
Cellulose acetate butyrate	Better orientation stability, chemical and moisture resistance, innately hydrophobic	Acetone: dimethyl acetamide (1:4 v/v)	Hydrophobic nature limits the application in the biomedical field	Oil absorption	[96]
Carboxymethyl cellulose	Enhanced biodegradability, Good interaction with carboxyl and amine groups in drugs	Distilled water	Rigid structure and high viscosity result in gel formation	Drug delivery, Food preservation	[97-99]
Hydroxy propyl cellulose	Better solubility in common solvents, especially water and methanol, constant fibers diameter	Chloroform and dimethyl formamide	Low mechanical strength	Transdermal drug delivery, sensors	[100, 101]

246

247 **2.2.2 Lignin**

248 Lignin is a highly preferred option for ES because of its low cost, less pollution, and availability
249 of renewable sources (the second most abundant biopolymer in nature behind cellulose). Lignin
250 is mainly derived from wood pulping byproducts as the secondary cell wall of higher plants
251 contains lignin [102]. It provides mechanical stability, the polysaccharide binding site, and aids
252 in resistance to pathogenic invasion [103]. Lignin is considered a phenolic biopolymer since
253 enzyme based polymerization of three primary phenolic alcohols (monolignols-p-coumaryl,
254 coniferyl, and sinapyl) produces plant lignin. Since lignin molecules are widely crosslinked with
255 one other and different polysaccharides, the "native structure" is still unknown as it cannot be
256 removed from plants in their natural state [103]. The significant properties of lignin include
257 biocompatibility, biodegradability, antioxidant, and more chemical resistance than other

258 cellulose-based biopolymers. These properties have considerable potential for biomedical
 259 applications [104]. However, the major issues regarding the ES of lignin are low solubility, poor
 260 mechanical strength, impurities during the extraction process, low interaction with other
 261 biopolymer systems, and less viscoelasticity for ES [12]. It is mostly ES with other biopolymers
 262 to overcome the nanofibers fabrication of lignin. Some of the major biopolymers blended with
 263 lignin and their solvent systems are listed in Table 4. The SEM images of various biopolymers
 264 blended with lignin are shown in Fig. S2 [12, 105].

265 **Table 4. List of commonly blended biopolymers with lignin for the electrospinning**

Source	Commonly blended biopolymers	Solvent system	Potential applications	References
Natural	Chitosan	Glacial acetic acid (1.5% V/V or 5% v/v)	Food packaging, Ultrafiltration	[106, 107]
	Cellulose	Trifluoroacetic acid: dichloromethane (1:1 w/w)	Packaging, Filtration	[108, 109]
Synthetic	Polylactic acid	1,1,1,3,3,3-hexafluoro-2-propanol	Tissue engineering	[110]
	Polyethylene oxide	Dimethyl formamide	Tissue engineering	[111]
	Polyvinyl alcohol	Distilled water	Antimicrobial, Drug delivery	[112, 113]

266

267 **2.2.3 Chitosan and Chitin**

268 Chitin and chitosan (CS) have a repeating structure of (1,4)-b-N-acetyl glycosaminoglycan and
 269 its deacetylate derivative, respectively [114]. The chemical structure of chitin is almost identical
 270 to that of cellulose except for the organic group R, which in chitin is NHAc rather than OH.
 271 Crab, shrimp, prawn, and insect exoskeletons, as well as the cell walls of mushrooms, include the

272 most readily accessible animal derived biopolymer, chitin [115]. Chitin is produced yearly at
273 1011 tons but is mainly discarded in the fishing industry. The chitin nanofibers had a 10–20 nm
274 diameter and a good aspect ratio [116]. The application of chitin is somewhat limited due to its
275 insoluble character in most solvents [117].

276 Chitin deacetylation produces the polysaccharide biopolymer chitosan. In CS, the R group is
277 NH_2 instead of OH in cellulose. Because D-glucosamine repeat unit C-2 possesses a $-\text{NH}_2$
278 functional group, chitosan is a better chelating agent than cellulose [118]. Deacetylated
279 polysaccharides are transformed into polyelectrolytes in an acidic medium ($\text{pH} < 6.5$) and are
280 more likely to dissolve because they lose their charges and precipitate due to amine ($-\text{NH}_2$) group
281 deprotonation [119]. Chitosan properties in a solution are generally determined by molecular
282 weight, deacetylation, polymer charge, ionic strength, and pH [120]. The most well-known
283 chitosan properties include biocompatibility, biodegradability, metal chelation,
284 mucoadhesiveness, and antibacterial [121, 122]. Owing to the mentioned properties, it is a
285 polymer of high importance in biomedical goods, cosmetics, chemicals, and medicines [123].

286 The primary issue with the CS is that it can only be dissolved in an acidic medium ($\text{pH} 2-6$) and
287 becomes polycationic, forming a viscous solution. So, high voltage is required for the ES of CS
288 due to high viscosity. Due to the application of high voltage, CS NFs generally show bead
289 formation [124-127]. The other issue includes the strong intramolecular hydrogen bonding
290 created in chitosan solutions since the polymer chain motion in response to the electric field is
291 usually suppressed, and for continuous fiber fabrication to occur during ES, it induces minimal
292 accessibility for the spinning of the polymer [128, 129]. These problems can be addressed by
293 employing solvents like trifluoroacetic acid (TFA), or a combination of TFA and
294 dichloromethane (DCM), or by adding additional solvents to the former mixture such as ethanol,

295 1,4-dioxane, or acetone [130, 131]. Chitosan solution in TFA generates soluble salt residues of -
296 $\text{NH}_3^+\text{CF}_3\text{COO}^-$ because the amino groups of CS form salts with TFA, disrupting the regular rigid
297 interaction between chitosan molecules and hence resulting in the dissolution of the polymer,
298 increasing the accessibility for ES and the charge density of CS solution [132]. Due to the high
299 volatility of the TFA solvent, the resulting NFs solidify rapidly. The fundamental issue with the
300 TFA based CS NFs is the low mechanical strength in the aqueous media which is the main
301 requirement for any biomedical application [133]. However, blending CS with other biopolymers
302 can efficiently address this issue. Some of the major blended biopolymers used with CS to
303 produce NFs and the solvent systems are listed in Table 5. The morphologies of various blended
304 chitosan nanofibers are shown in Fig. S3 [17, 134].

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316 **Table 5. List of various commonly blended biopolymers for the electrospinning of chitosan**

Source	Commonly blended biopolymers	Solvent system	Application	References
Natural	Alginate	Ethanol in water (20% v/v) OR Acetone: dimethyl acetamide (2:1 w/w)	Wound dressing, Skin repair, Drug delivery	[135-137]
	Pullulan	Aqueous acetic acid (50% v/v)	Drug delivery, wound dressing	[138, 139]
	Cellulose	Trifluoro acetic acid: acetic acid (7:3 w/w)	Adsorption, Food packaging	[133, 140]
	Hyaluronic acid	Distilled water	Drug delivery, wound dressing	[11, 67]
Synthetic	Polyethylene oxide	Acetic acid (various percentages)	Tissue engineering, Sensors, wound healing	[141-143]
	Polyvinyl alcohol	Acetic acid (various percentages) OR Distilled water	Metal ion removal, Drug delivery	[144, 145]
	Polylactic acid	Trifluoro acetic acid: dichloromethane (70:30 v/v)	Water filter, Wound dressing, Drug delivery	[146-148]

317 **2.2.4 Hyaluronic Acid**

318 Natural Hyaluronic Acid (HA) is a linear polysaccharide composed of alternating disaccharide
319 units of D-Glucuronic acid and N-acetyl D-glucosamine. Its molecular weight ranges from 100
320 to 8,000 kDa depending upon the source [149]. The HA molecular weight is essential in several
321 biological activities, as shown in Table 6. As a component of most mammalian connective
322 tissues, HA helps to maintain structural integrity and viscoelasticity. HA's primary distribution in
323 connective tissues and important role in cell adhesion, proliferation, migration, and granulation
324 tissue development are all HA's biological features [150].

325

326 **Table 6. Correlation of hyaluronic acid molecular weight with biomedical applications**

Molecular weight (Da)	Applications	References
6×10^6	Skin inflammation treatment	[151]
$>1 \times 10^6$	Joint inflammation treatment	[152, 153]
1.2×10^5	Cancer treatment	[154, 155]
2×10^4	Immunomodulation	[156]
1×10^6	Removal of reactive oxygen species by UV-rays	[157, 158]
1×10^6	Epidermal cells growth regulation	[159]
$<1 \times 10^4$	Skin metabolism regulation and antiaging	[160, 161]
$2-3 \times 10^3$	Promoting collagen synthesis	[162, 163]
$<1 \times 10^4$	Joint health improvement by oral formulation	[164]

327

328 The most distinguishing features of HA are its biodegradability, biocompatibility, and negligible
329 induction of immunogenic response, which makes HA an important biopolymer for various
330 biomedical applications. The half-life of HA depends upon the location of the organs. It has a
331 short half-life of 24 hours in skin tissues and is around 70 days long in some eye regions (up to
332 70 days) [165]. Injectable HA gels and solutions are used as dermal fillers to restore skin volume
333 and reduce wrinkles' appearance. HA is also used in other cosmetics because of its safety and
334 effectiveness. Drug delivery [166], osteoarthritis [167], and cosmetics [168] are only a few of the
335 medical uses of HA. Furthermore, HA is often recognized as the most hydrophilic, naturally
336 occurring polymer. The strong absorption characteristics of HA for wound exudates make it a
337 preferred option for wound dressings [169].

338 The major issue with HA is the low mechanical strength in physiological conditions. HA is
339 easily depolymerized invitro by acids and oxygen in the presence of thiols and ferrous based

340 reducing agents. [170]. Additionally, enzymatic or reactive species-mediated HA degradation
341 may also occur in vivo and limit the biomedical applications of HA. Since HA is only soluble in
342 the aqueous based media but even at low concentrations, the electrospinnability is poor due to
343 the high viscosity and surface tension due to its ionic nature. Counter-ions in aqueous solutions
344 also increase HA solutions' viscosity [171]. Due to the high viscosity, HA polymer chains could
345 not interact with one another, contributing to jets' instability and the subsequent creation of
346 discontinuous and heterogeneous NFs. Due to aqueous based solvents, the jet does not evaporate
347 between the collector and the needle, leading to droplets or clogging in the needle. HA's
348 relatively high surface tension requires a high voltage to generate more intense electrostatic
349 forces to fabricate NFs, resulting in beading. Therefore, the high surface tension of HA must be
350 decreased for the fabrication of continuous NFs [172]

351 To address these mentioned issues, the first approach is to blend HA with other biopolymers for
352 the NFs fabrication [173]. Major biopolymers used for the ES of HA are listed in Table 5. The
353 second approach uses solvents that disrupt the H-bonding in the HA chains [172]. That approach
354 increases the flexibility of the chains, but the downside is the addition of toxic materials that
355 reduce the potential for biomedical applications [174]. The third approach uses a surfactant to
356 reduce surface tension [175]. The fourth approach is the modification of HA, which can be
357 physical or chemical crosslinking. Conventional crosslinking is mainly accomplished by using
358 aldehydes, hydrazides, or sulfides. However, the downside of the chemical crosslinking is the
359 increase in cytotoxicity of the resultant NFs. Recently, 1-ethyl-3-(3-dimethylaminoisopropyl)
360 carbodiimide modification has been used and considered better than the conventional
361 crosslinking as the latter result in a non-cytotoxic product [175]. The physical modification is
362 mostly done using silver nanoparticles to develop self-crosslinking [176]. Some of the major

363 biopolymers used with HA to produce NFs and the solvent systems are listed in Table 7. Some
 364 important HA blended nanofibers morphologies are shown in Fig. S4 [11, 177].

365 **Table 7. List of various biopolymers blended with hyaluronic acid for electrospinning**

Source	Commonly blended biopolymers	Solvent system	Application	Reference
Natural	Cellulose	Distilled water	Therapeutic delivery	[178]
	Collagen	Hexafluoro isopropanol	Wound healing, Tissue engineering	[179, 180]
Synthetic	Polyvinyl alcohol	ethanol: water: benzyl alcohol (50:40:10 v/v)	Wound healing, drug delivery	[175, 181, 182]
	Polyethylene oxide	Water and 2,2,2-Trifluoroethanol: ethanol (1:10)	Drug delivery, Wound healing	[183, 184]
	Polycaprolactone	Ethanol: chloroform (7:3) OR formic acid: acetic acid (7:3)	Drug delivery, Wound healing, Tissue engineering	[185, 186]

366

367 **2.2.5 Alginate**

368 Alginate (AG) is a linear anionic polysaccharide having mannuronic acid and glucuronic acid,
 369 referred to as M-blocks and G-blocks, respectively, and (1,4)-glycosidic bonds connect both
 370 blocks. AG is mainly obtained from bacteria and algae, with algae being the most common
 371 source [187]. AG has a similar role to cellulose in plants by giving strength and flexibility to the
 372 algae. Some bacteria (such as Azotobacteria and Pseudomonas) may also make AG [188].
 373 Several features of AG vary depending on the source. Due to their high G-block concentration,
 374 seaweed AG is often employed for biomedical applications [189]. In contrast, bacterial source-
 375 based AG has high M-blocks concentration and often demonstrates enhanced immunogenicity
 376 and cytokine production [190].

377 AG is an appealing material for increasing production and industrial applications since it is
378 readily available, economically viable, and ecologically beneficial. AG is also nontoxic when
379 employed in medical applications and has a low immunogenetic response in people because it
380 has excellent absorptivity, biocompatibility, biodegradability, and nontoxicity [191]. Despite
381 numerous beneficial characteristics, AG is difficult to electrospun, and pure AG electrospinning
382 in organic or aqueous solvents has not been reported. Some of the properties of AG that have
383 been recognized for its poor electrospinnability include its polyelectrolyte nature, stiff and
384 prolonged structure, a tendency to form a gel at low concentration, high electrical conductivity,
385 and high surface tension [191-193]. These problems are mainly addressed using a cosolvent
386 system, blending with other biopolymers, surfactants, and chemical modifications [194]. The co-
387 spinning polymers with better electrospinnability when blended with AG result in enhanced AG
388 electrospinnability. The blending with other biopolymers boost the electrospinnability by
389 producing charge repulsion in polymer chains of AG hence improving the flexibility of AG and
390 generating new hydrogen bonds [191]. Some of the significant blended biopolymers and solvent
391 systems of AG are listed in Table 8, and some major nanofiber morphologies are shown in Fig.
392 S5 [14, 195].

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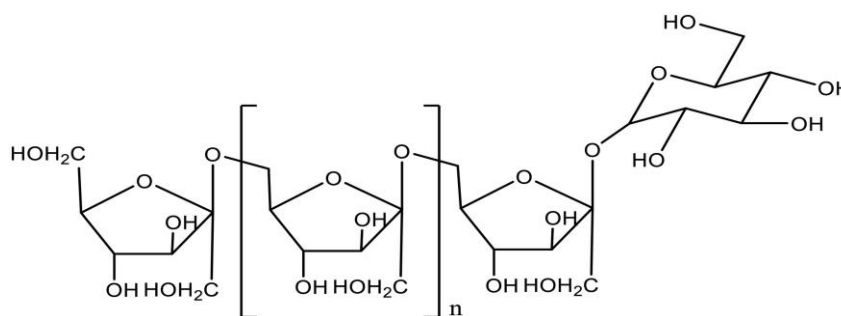
399 **Table 8. List of various biopolymers blended with alginate for the electrospinning**

Source	Commonly blended biopolymers	Solvent system	Application	Reference
Natural	Gelatin	Distilled water	Cell culture, Wound healing, dye adsorption	[196-198]
	Pullulan	Distilled water	Food packaging	[199]
Synthetic	Polycaprolactone	Aqueous acetic acid	Cell culture, Drug delivery, Wound healing	[200, 201]
	Polyethylene oxide	Distilled water	Wound dressing, Dye adsorption	[202-204]
	Polyvinyl alcohol	Acetone: dimethyl formamide (2:3 v/v)	Tissue engineering, Sensor	[205-207]

400

401 **2.2.6 Levan**

402 For years, scientists have been intrigued by levan, a fructan homopolysaccharide with
 403 amphiphilic properties [208]. The structure consists of residues of D-fructo-furanose are held
 404 together by 2,6-glycosidic linkages and is synthesized by a wide range of microorganisms
 405 (including Acetobacter, Bacillus, Halomonas, Microbacterium, and Streptococcus) and some
 406 plants [209] (Fig. 9).

407 **Fig. 9 Chemical structure of levan [1]**

408 Any enzymes do not digest Levan once it enters the body since it does not affect the
409 physiological environment of animals, unlike other polysaccharides like cellulose and starch
410 [210]. This property can make this material an ideal candidate for drug delivery applications. In
411 addition, they are intrinsically mucoadhesive and can be used in tissue engineering [1]. The main
412 issue with levan ES is its inconsistent fluid properties at low or high concentrations. Levan
413 displays Newtonian behavior for concentrations of up to 30% by mass, and between 30% and
414 55%, it shows shear thinning behavior, and for greater than 55% by weight, no plateau at low
415 shear rates is obtained [211]. However, its nanofibers have been reported at 60% weight
416 concentration in water [212]. Therefore, this material requires further studies to acknowledge its
417 ES potential and to find better biopolymer for blending to enhance its natural properties and
418 overcome its drawbacks.

419 **2.2.7 Kefiran**

420 Kefir is a probiotic and jelly-like grains complex of bacteria and yeasts that produces kefiran, a
421 glucogalactan heteropolysaccharide copolymer of D-glucose and D-galactose [213] (Fig. S6).
422 Kefiran has long been recognized as a possible biomaterial. It was electrospun for the first time
423 in 2014 from a water-based solution with no additives. Since then, various efforts have been
424 made to produce multifunctional nanofibrous scaffolds based on kefiran [214, 215].

425 Kefiran has anti-inflammatory, antibacterial, antifungal, and excellent biocompatibility
426 properties. Probiotic properties have been proven to stimulate the development of healthy
427 microflora while inhibiting the growth of cancer cells and bacteria. The best advantage of kefiran
428 is its solubility in water and other commonly available solvents [216]. However, additional study
429 is needed from the ES perspective, as kefiran is expected to generate a new, fascinating category
430 of biomaterials with enhanced properties. Some recent biopolymers blended with kefiran and the

431 solvent system for ES are listed in Table 9, and some major nanofiber morphologies are shown
 432 in Fig. S7 [18].

433 **Table 9. List of various biopolymers blended with kefir for the electrospinning**

Source	Commonly blended biopolymers	Solvent system	Potential applications	References
Natural	None	Deionized water	Drug delivery	[18]
	Chitosan	Aqueous acetic acid (2% v/v)	Tissue engineering	[217]
Synthetic	Polyvinyl alcohol	Distilled water	Yet to be reported	[218]
	Polyethylene oxide	Distilled water	Food packaging	[215]

434

435 Fig. 10 shows the SEM images of major polysaccharides nanofibers with different biopolymers
 436 and their ratios.

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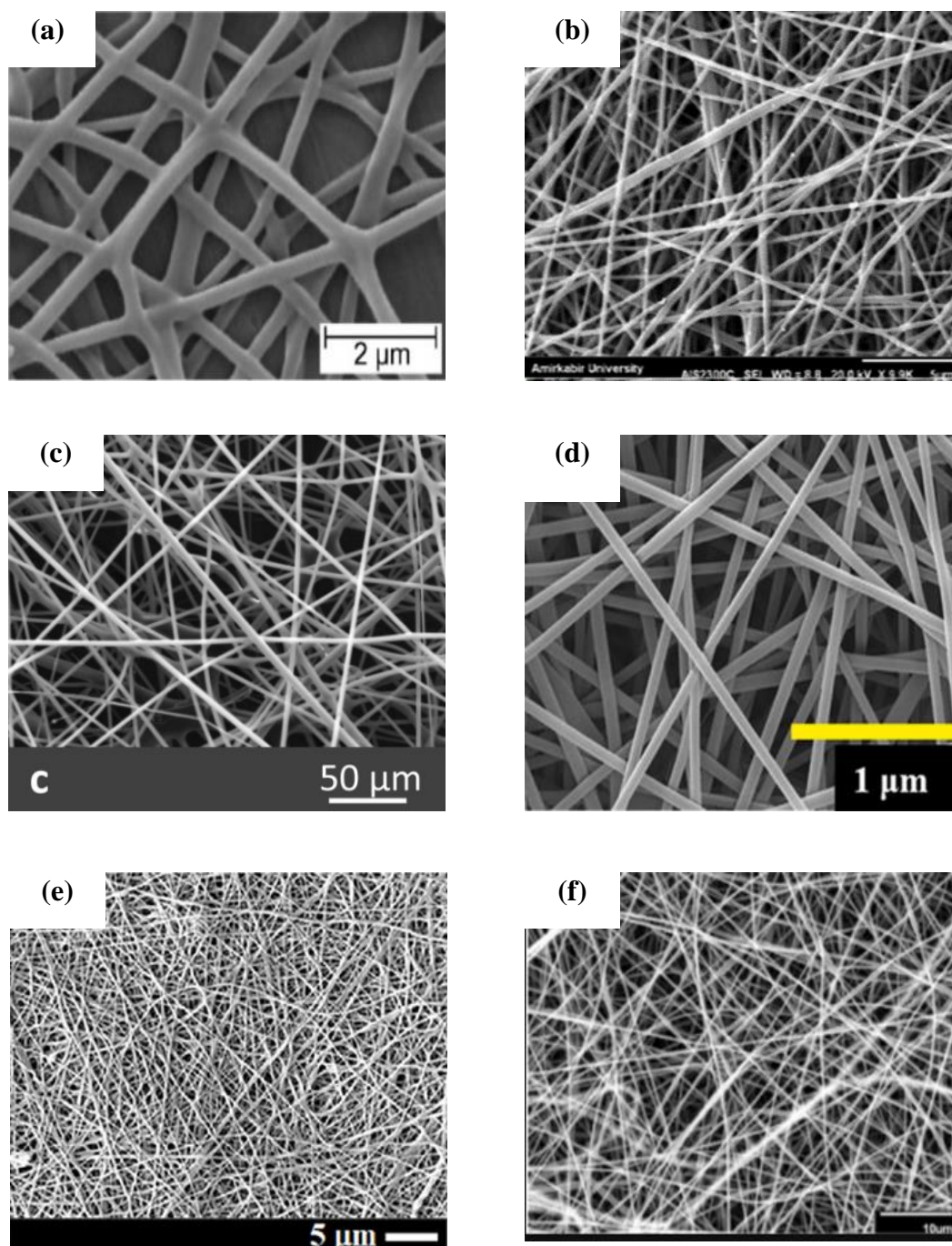


Fig. 10 SEM images of major polysaccharides nanofibers with other biopolymers, (a) Carboxymethyl cellulose: polyvinyl alcohol: (2:10 % w/v) [8] , (b) Hyaluronic acid: polyvinyl alcohol (1:2) [11], (c) Lignin: polyethylene oxide (97:3) [12], (d) Alginate: polyvinyl alcohol (8:2) [14], (e) Chitosan: alginate (1:1) [17], (f) Pure kefirin 4% [18]

443 2.2.8 Collagen and Gelatin

444 Collagen is the most common protein in mammals and makes up the ECM of numerous
 445 mammalian organs and tissues, including skin, bone, blood vessels, tendons, and ligaments
 446 [219]. Crosslinked tropocollagen units comprise three polypeptide chains that form a right-
 447 handed triple helix stabilized by interstrand hydrogen bonding and intrastrand interaction.
 448 Collagen protects the ECM's biological and structural integrity because of its high tensile
 449 strength and mechanical resilience [220]. Only one of the 29 collagen variations, collagen I, is
 450 used to produce collagen-based biomaterials. Because of its biodegradability, low antigenicity,
 451 customizable mechanical properties, ability to interact with a range of cell types, and capacity to
 452 form three-dimensional scaffolds, collagen is a material of interest in tissue engineering and
 453 therapeutic applications [221]. The major problems associated with collagen ES include loss of
 454 physiochemical structure due to high voltage, no swelling in the water; instead, it is dissolved
 455 instantly, and low denaturation temperature [222]. These shortcomings have been overcome by
 456 blending with other biopolymers (Table 10) and by using modifications in the structure [222]

457 **Table 10. List of various biopolymers blended with gelatin for the electrospinning**

Source	Commonly blended biopolymers	Solvent system	Potential applications	Reference
Natural	Chitosan	Distilled water	Tissue engineering, Wound dressing	[223, 224]
	Hyaluronic acid	Aqueous acetic acid	Tissue engineering, Wound healing	[179, 180, 225]
Synthetic	Polycaprolactone		Ulcer treatment, Wound healing, Tissue engineering	[226-228]
	Polylactic acid	Ethanol (80% v/v)	Tissue engineering,	[229, 230]
	Polyvinyl alcohol	Aqueous acetic acid (2% v/v)	Drug delivery, Wound dressing	[231, 232]

458 When collagen is hydrolyzed, it loses its native structure and transforms into gelatin. Two major
459 events occur of this loss: breaking the intermolecular glycosylation linkage between lysine and
460 hydroxylysine and breaking intramolecular disulfide bridges [233]. Collagen and gelatin are
461 biopolymers with a fundamental structure composed of as many as 20 amino acids in varying
462 quantities, depending on the hydrolysis procedure used to convert collagen to gelatin. However,
463 there is a three amino acid sequence of L-arginine-glycine-aspartic acid (RGD) which is
464 recognized by integrins to facilitate cell attachment.

465 The major features of gelatin include non-immunogenicity, biocompatibility, biodegradability,
466 adaptability, cheap cost, and no denaturation phenomena. Thus, gelatin is used in many ways in
467 the food and biomedical sectors, especially in tissue engineering. The major plus point is that
468 after being subjected to ES, it has no denaturation phenomena and is preferred over collagen.
469 The major drawback for gelatin NFs is the dissolution in the aqueous conditions. Gelatin is
470 usually crosslinked by several crosslinkers, mostly aldehydes, including glutaraldehyde (GA),
471 carbodiimides, succinimide, and genipin (GEN), for resolving dissolution issues. GA and GEN
472 are considered the most suitable crosslinking agents because of their low toxicity and resistance
473 to degradation in cell culture conditions at 37 °C [234].

474 Collagen and gelatin are usually methacrylate modified and resulting in ColMA and GelMA,
475 respectively. ColMA has been widely researched for biological uses in the form of hydrogels
476 [235], but its usage in ES has been restricted due to structural denaturation issues. Since ES
477 produces GelMA rather than ColMA, it is a common electrospinnable material because of its
478 high biocompatibility and morphological and chemical resemblance to ECM [236]. Furthermore,
479 it promotes neovascularization and water adsorption, and its biodegradation rate is rapid and

480 simple to control. It has been widely used in tissue engineering [237]. Fig. S8 shows some
481 essential collagen and gelatin nanofibers with commonly blended biopolymers [10, 238].

482 **2.2.9 Elastin**

483 Elastin is a naturally occurring protein with natural fiber morphology and is majorly found in
484 animal ECM. This insoluble, elastomeric protein is formed by crosslinking tropoelastin, the
485 protein's precursor (Fig. S9) [239] . Elastin has remarkable qualities, including a half-life of more
486 than 70 years, outstanding durability, reversible stretchability, biocompatibility, and resistance to
487 chemical and mechanical impacts. Elastin fibers have Young's modulus between 300 and 600
488 kPa and can be stretched up to 220% of their original length. They can withstand billions of
489 stretch and recoil cycles without breaking [240].

490 Elastin found its application in wound healing and tissue engineering due to the mentioned
491 intrinsic properties [241]. Since elastin is both resistant and insoluble, it presents specific
492 difficulties as a starting material for ES. However, the solubility of elastin may be increased by
493 acidic or basic hydrolysis. These preparations have several drawbacks, including compositional
494 variability, potential loss of cellular signaling capabilities, and the need for crosslinking by
495 glutaraldehyde to increase the water stability of the fiber mats. It is frequently mixed with
496 additional polymers such as tropoelastin or elastin-like recombinamers (ELRs) 200 for rendering
497 pure elastin spinnable. Major biopolymers blended with elastin are enlisted in Table 11, and
498 some morphologies are shown in Fig. S10 [13, 242].

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502 **Table 11. List of various biopolymers blended with elastin for the electrospinning**

Source	Commonly blended biopolymers	Solvent system	Potential application	References
Natural	Alginate	Deionized water and hexafluoro isopropanol	Tissue engineering	[34]
	Collagen	Deionized water	Tissue engineering	[243, 244]
	Hyaluronic acid	Hexafluoro isopropanol	Tissue engineering	[225, 245]
	Chitosan	Trifluoroacetic acid: dichloromethane (7:3)	Tissue engineering, Wound healing	[13, 246, 247]
Synthetic	Polylactic acid	Deionized water and dichloromethane	Tissue engineering, Wound healing	[248, 249]

503

504 **2.2.10 Silk fibroin**

505 Silk fibroin (SF) is a fibrous protein from silkworm larval cocoons, often *Bombyx mori* or
506 *Antheraea assama*. It consists of polypeptide chains ranging in size from 200 to 350 kDa, and
507 each chain consists of a tandem repeat of hydrophobic heavy and hydrophilic light chains held
508 together by disulfide bonds [250]. As a result, a semi-crystalline fishnet structure is formed, with
509 the crystalline regions acting as pressure sponges that disperse that force evenly over the whole
510 fibroin network [247, 248]

511 Due to its accessibility, cheap cost, cytocompatibility, bioactivity, biodegradability,
512 thermostability, excellent mechanical qualities, and minimal immunogenicity, SF has seen an
513 uptick in usage as a biomaterial for tissue engineering and various biomedical applications in the
514 previous decade [251, 252]. The solution is prepared in formic acid or hexafluoro-2-propanol for
515 the ES of pure SF [253]. Organic solvents like methanol are required to stimulate the production

516 of a more stable antiparallel-sheet conformation (silk II) in the NFs. There is a risk that organic
 517 and caustic solvents can alter the structure and bioactivity of biomolecules, particularly sensitive
 518 therapeutics in biomedical applications [254]. As a result, it is recommended to use less severe
 519 conditions and refrain from employing organic solvents to reduce their toxicity. Mostly SF is
 520 also electrospun with other biopolymers to overcome the ES issues. Some of the most common
 521 blended biopolymers are listed in Table 12. The effect of the solvent system on the SF can be
 522 seen in Fig. 11; aqueous media results in uniform morphology, while the organic solvent
 523 hexafluoro isopropanol results in a wide range of diameter. Overall, SF seems promising for
 524 developing various tissue engineering materials, especially for treating burnt or infected wounds.

525 **Table 12. List of various biopolymers blended with silk fibroin for the electrospinning**

Source	Commonly blended biopolymers	Solvent system	Potential applications	References
Natural	Collagen	Aqueous acetic acid	Tissue engineering	[255, 256]
	Hyaluronic acid	Deionized water	Tissue engineering, Wound healing	[257]
Synthetic	Polyethylene oxide	Deionized water	Tissue engineering, Wound healing	[258, 259]
	Polyvinyl alcohol	Aqueous acetic acid (90% v/v)	Tissue engineering, Drug delivery, Wound dressing	[260-263]

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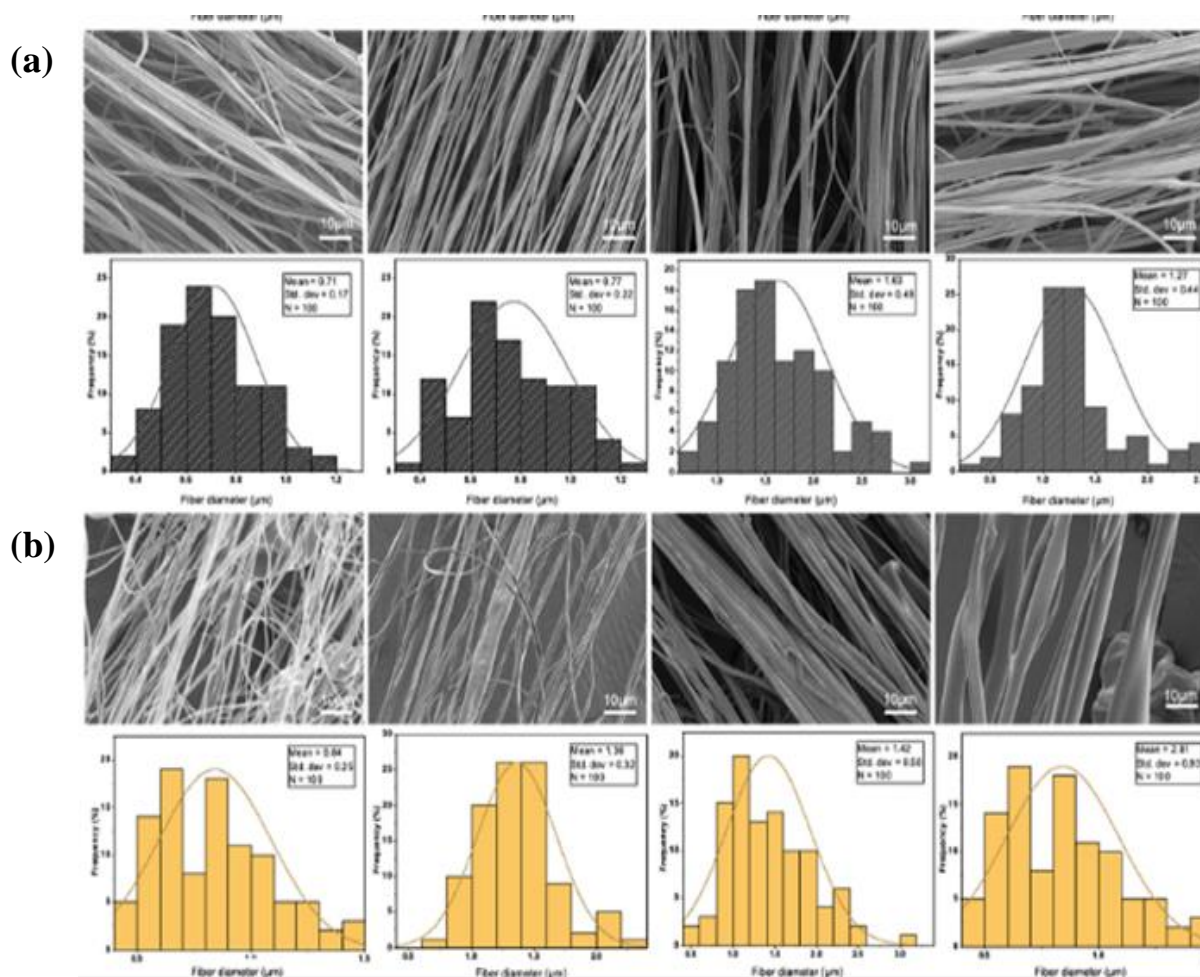


Fig. 11 SEM images of silk fibroin nanofibers with other biopolymers, (a) Polyethyleneimine: silk fibroin (10:90) in aqueous media, (b) Silk fibroin (8% w/v) in organic solvent (hexafluoroisopropanol) [9]

533 Fig. 12 shows the SEM images of major proteins nanofibers with different biopolymers and their
 534 ratios.

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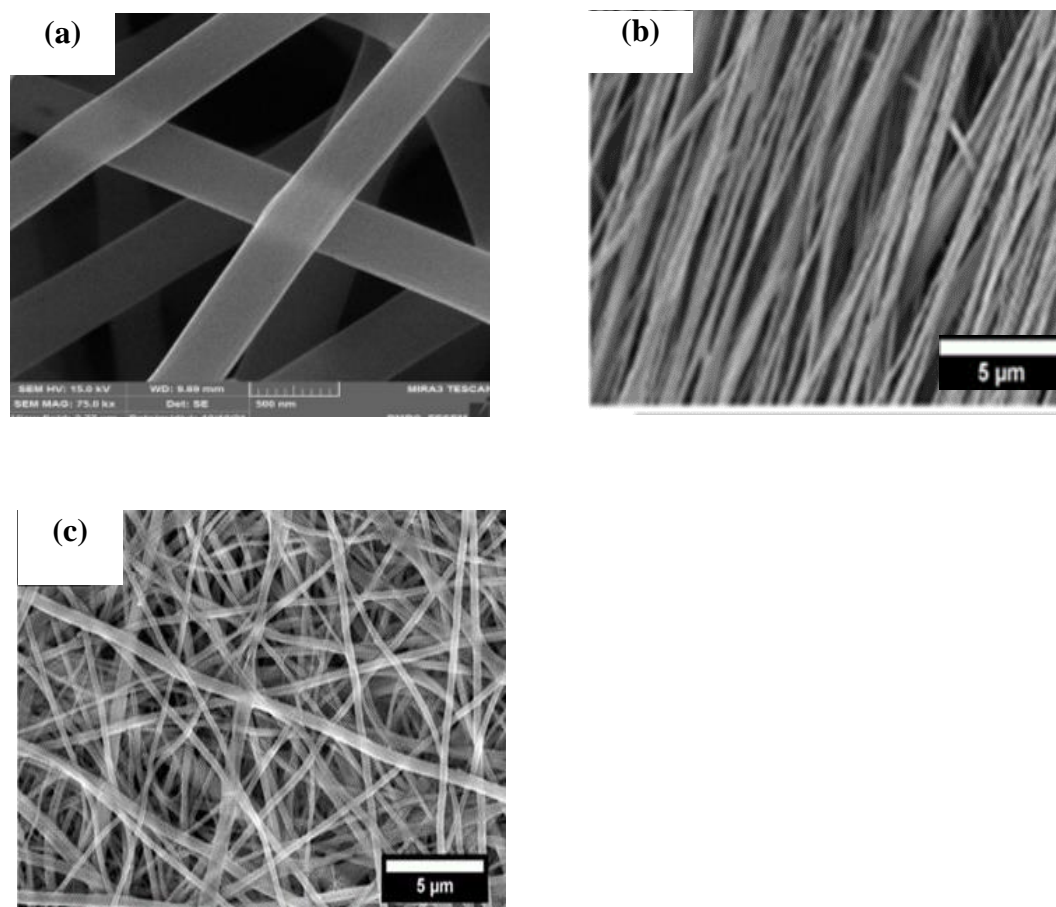


Fig. 12 SEM images of major proteins nanofibers with other biopolymers, (a) Gelatin: Alginate (1:1) [10], (b) Silk fibroin: polyethyleneimine (90:10) in aqueous media [9], (c) Elastin: chitosan (5.5:4 w/v) [13]

539

540 **3. Trends in biopolymers electrospinning**

541 We use the ScienceDirect database to study the current trends and progress for biopolymeric
 542 nanofibers (Fig. 13). Biopolymers were divided into two groups of polysaccharides and proteins,
 543 depending on their biochemical nature. In the case of the polysaccharides group 212% increase,
 544 while in the case of proteins, a 152% increase is seen in the nanofibers research from 2018 to
 545 2022. For polysaccharides, cellulose is the top-researched biopolymer, while collagen and gelatin
 546 have been the top research biopolymers since 2018. However, in our review, we have

547 highlighted polysaccharides newly introduced in ES. Kefir and levan have been gaining interest
 548 for the past few years, but still, much study is required to unveil their potential.

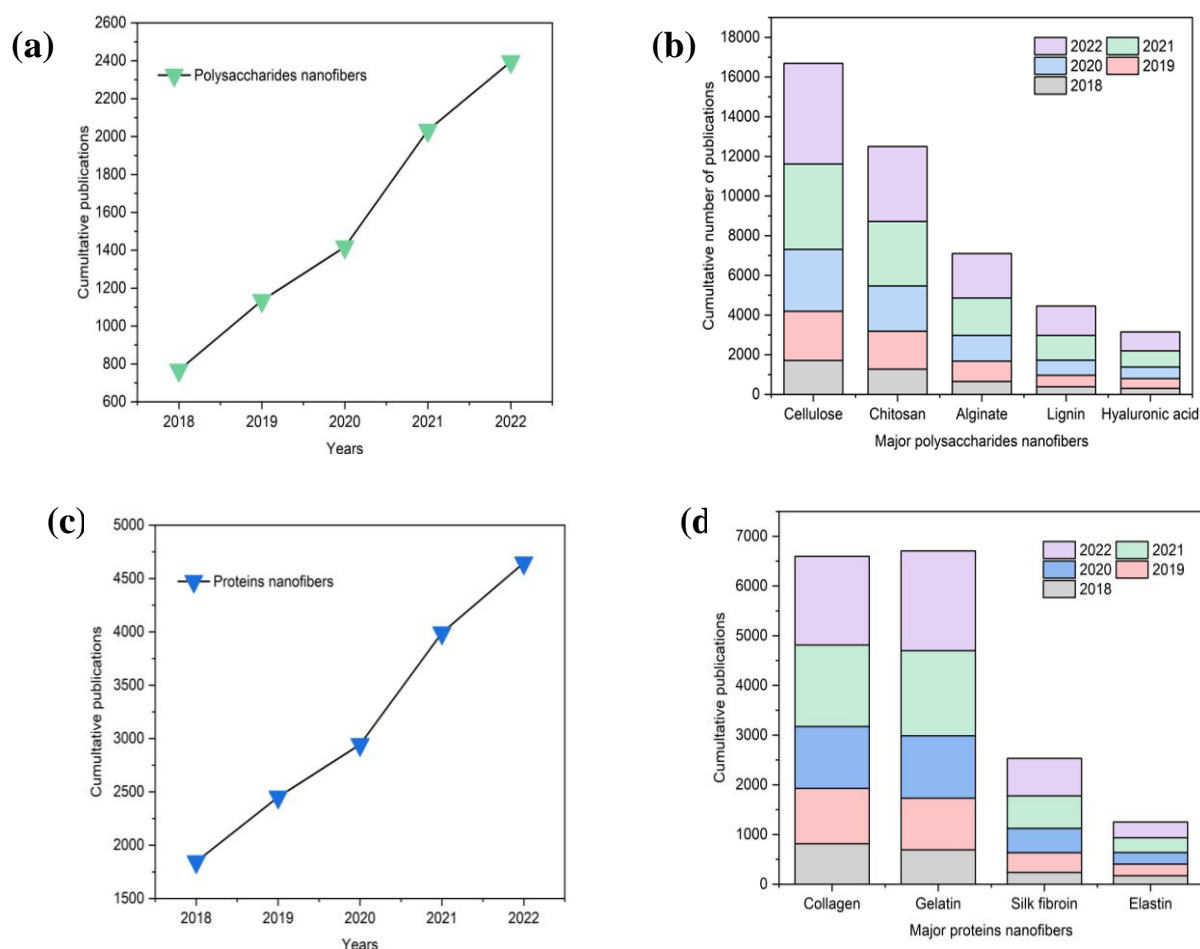


Fig. 13 Cumulative number of publications in ScienceDirect platform in the last five years (2018-2022), (a) Total polysaccharides based nanofibers, (b) Major polysaccharides nanofibers, (c) Total protein based nanofibers, (d) Major proteins nanofibers

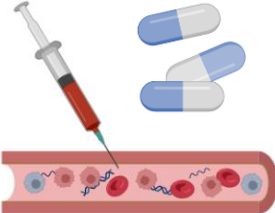

549 **4. Commercial products**

550 Much study has been dedicated to the fast expanding field of electrospinning. Upscaling to
 551 industrial standards and commercialization are the ultimate goals of every research field.
 552 Regarding electrospinning, however, repeatability issues and batch-to-batch variability are the
 553 biggest problems across all methods [264]. This is a major hindrance to the widespread

554 commercialization and industrialization of electrospinning, particularly for use in the medical
555 field. Table 13 details several cutting-edge electrospinning-based medicinal supplies.

556

557 Table 13. Recent applications of electrospinning methods for biopolymers nanofibers based wound healing system

Product category	Brand name	Manufacturer	Main component	Key features	References
Drug delivery patches 	Rivelin® patch	Bioinicia (Valencia, Spain)	---	The system is specially designed for mucosal surface unidirectional drug delivery.	[265]
	Zeus Bioweb™	Zeus Industrial Products, Inc. (Orangeburg SC, USA)	Polytetrafluoroethylene	Ultrasmall fibers with the least chemical reactivity	[266]
Surgical sutures and wound healing dressings 	ReDura™	MEDPRIN (Guangzhou, China)	Polylactic acid	Material is similar to extracellular matrix (ECM) and promotes rapid repair and regeneration.	[267]
	ReBOSSIS®	Ortho ReBirth (Yokohama-shi Kanagawa pref., Japan)	TCP (β-Tricalcium Phosphate), Bioabsorbable Polymer and SiV (Silicone-containing Calcium Carbonate)	Bone filler that promotes the bone formation	[268]
	HealSmart™	PolyRemedy®, Inc. (Concord, MA, USA)	Hyaluronic acid	Antimicrobial Dressings	[269]
	PK Papyrus®	Biotronic (Berlin, Germany)	Polyurethane	Thin and elastic membrane for stent coating	[270]
	Surgiclot®	St. Theresa Medical Inc. (Eagan, USA)	Dextrin and fibrin	Sealant for the bone bleeding	[271]

558 **5. Future perspectives**

559 Despite widespread agreement that electrospinning is a fascinating method with promising
560 applications, issues still need to be addressed. Various hurdles to fabricating these biopolymeric
561 nanofibers include innate poor electrospinnability of biopolymers, lack of individual
562 biopolymers properties to accomplish the applications solely, and optimizing the solvent system
563 for the nanofibers fabrication. This review focusses on all three mentioned issues and addresses
564 the solutions for individual biopolymers. The blending of biopolymers can solve the lack of
565 properties issue. The major results of this study suggested that the functionalization of
566 biopolymeric materials and then the blending with other biopolymers is crucial for developing
567 and improving these materials for use in targeted applications. However, the major concern is the
568 insolubility of biopolymers and the optimization of the solvent system. The solvent system plays
569 a critical role in the fabrication of the nanofibers. This study provides insight into the current
570 solvents systems for the major biopolymers nanofibers fabrication.

571 However, it was observed that most of the solvents employed for the fabrication are organic
572 solvents, and they are a concern from the environmental and the user's point of view. During the
573 ES procedure, most of the solvents are evaporated; however, some traces are still left in the
574 fabricated nanofibers, resulting in the commercial level pharmaceutical development of
575 biopolymeric nanofibers. Melt electrospinning might seem like a straightforward way to
576 overcome the organic solvent based procedures. However, the process has drawbacks regarding
577 the fibers' complexity, fibers with a large diameter, polymer-related thermal degradation, and
578 incompatibility with several high throughputs. On the other hand, needleless electrospinning is
579 considered the most advance electrospinning method, but in this technique, a large liquid surface
580 that is exposed to the air and evaporates extremely volatile solvents into the environment results

581 in raised concerns for the user and the developed product. To overcome the solvent related
582 issues, researchers should develop "green" solvents as the alternative to the harmful toxic
583 solvents whose residues will not be harmful or will be affecting the biomedical applications and,
584 above all, will increase the electrospinnability of the blend solution.

585 The other major issue noted in the research is the lack of generalized solvent-based models for
586 the ES process biopolymers. Even though stimulation models have been studied, no one has yet
587 created a model that can reliably forecast the needle-based or needleless electrospinning
588 parameters. Therefore, most ES experiments depend on parametric analyses and empirical
589 knowledge of the process requirements. The current literature lacks studies on developing
590 standard protocols to make the fabrication more standardized rather than focusing on just
591 changing the process parameters (voltage, needle distance, and flow rate). To overcome these
592 restrictions, scientists should be more forthcoming with their findings on optimizing the solvent
593 systems and how the solvent system will affect different properties of biopolymeric under the
594 same electrospinning parameters and vice versa. In addition, more focus should be put on
595 developing green solvents for biopolymeric nanofibers to replace harmful and toxic solvents.

596 **6. Conclusion**

597 Biopolymers are the materials in focus to replace petroleum based polymers. Various
598 biopolymers have proved to be perfect candidates in different industries, especially in the health
599 sector, due to their natural features like biocompatibility, antimicrobial activity, and
600 biodegradability. The fibrous morphology of biopolymers is essentially required for effective
601 biomedical applications, including tissue engineering, drug delivery and wound healing.

602 Electrospinning (ES) has emerged as the most productive and modern technique for fabricating
603 biopolymeric nanofibers. ES provides the flexibility of combining various biopolymers without

604 toxic crosslinkers, which is nearly impossible with other conventional techniques. The resultant
605 nanofibrous morphology provides better surface area, mechanical properties, and compatibility.
606 However, electrospinning of biopolymers is still going through the developmental stages, and
607 there are many issues, including the use of toxic solvent systems, no focus on parameter based
608 studies in the current literature and the lack of reproducibility. The solvent system issue can be
609 resolved by formulating environmental friendly green solvent systems. For the other issues,
610 more parametric studies are required to make the ES procedure universal and user-friendly to
611 reduce variation among the batches. The focus on these recommendations can lead to better
612 utilization of the ES technique and increased commercialization of biopolymeric nanofibrous
613 products.

614 **Declaration of Competing Interest**

615 The authors state that no potential conflicts of interest or personal ties might have influenced the
616 work reported in this research.

617 **Declaration of generative AI in scientific writing**

618 The authors state that no AI software or AI based technologies were used for the writing
619 purposes of this review article.

620 **Authors contribution**

621 All authors contributed towards drafting and critically revising the paper and agree to be
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624

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