

Meta analaysis

Global patterns of insect herbivory in gap and understorey environments, and their implications for woody plant carbon storage

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Subject Editor: Lonnie Aarssen Editor-in-Chief: Dries Bonte Accepted 1 December 2017 Insect herbivory is thought to favour carbon allocation to storage in juveniles of shade-tolerant trees. This argument assumes that insect herbivory in the understorey is sufficiently intense as to select for storage; however, understoreys might be less attractive to insect herbivores than canopy gaps, because of low resource availability and – at temperate latitudes – low temperatures. Although empirical studies show that shade-tolerant species in tropical forests do allocate more photosynthate to storage than their light-demanding associates, the same pattern has not been consistently observed in temperate forests. Does this reflect a latitudinal trend in the relative activity of insect herbivory in gap versus understorey environments? To date there has been no global review of the effect of light environment on insect herbivory in forests. We postulated that if temperature is the primary factor limiting insect herbivory, the effect of gaps on rates of insect herbivory should be more evident in temperate than in tropical forests; due to low growing season temperatures in the oceanic temperate forests of the Southern Hemisphere, the effect of gaps on insect herbivory rates should in turn be stronger there than in the more continental temperate climates of the Northern Hemisphere. We examined global patterns of insect herbivory in gaps versus understories through meta-analysis of 87 conspecific comparisons of leaf damage in contrasting light environments. Overall, insect herbivory in gaps was significantly higher than in the understorey; insect herbivory was 50% higher in gaps than in understoreys of tropical forests but did not differ significantly between gaps and understories in temperate forests of either hemisphere. Results are consistent with the idea that low resource availability - and not temperature - limits insect herbivore activity in forest understoreys, especially in the tropics, and suggest the selective influence of insect herbivory on late-successional tree species may have been over-estimated.

Keywords: shade tolerance, herbivory tolerance, meta-analysis, forest dynamics, latitudinal gradient



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Introduction

Insect herbivory is thought to exert important selective pressures on the traits of late-successional tree species. A carbon conservation strategy of allocation to storage and defences at the expense of growth is considered critical for low-light survival (Coley et al. 1985, Myers and Kitajima 2007, Poorter et al. 2010): to persist in the understorey, plants must be able to recover from herbivory through mobilization of stored carbohydrates (Kitajima 1994, Kobe 1997, Walters and Reich 1999). This strategy is also known as herbivory tolerance, and it is assumed to relate to heavy herbivory pressure and severe damage (Rosenthal and Kotanen 1994, Rosenthal and Welter 1995, Tiffin and Rausher 1999). Thus, the view that conservative carbon allocation is fundamental for low-light success assumes that understorey insect herbivory is high enough as to select for herbivory tolerance.

Although some studies have found evidence linking seed-ling shade tolerance to carbon allocation to storage (Canham et al. 1999, Kitajima 1994, Poorter and Kitajima 2007), several others did not (Canham et al. 1999, Lusk and Piper 2007, Piper 2015, Piper et al. 2009, 2017). One possible explanation for the discrepancy is that, in some regions, the levels of understorey herbivory are insufficient to select for herbivory tolerance. Although many studies have examined the influence of light environment on insect herbivory (Niesenbaum 1992, Niesenbaum et al. 2006, Guerra et al. 2010, Salgado-Luarte and Gianoli 2010), at present we lack an overview of levels of insect herbivory damage in contrasting light environments at regional and global scales. Such an assessment would improve our understanding of selective pressures on plant traits in relation to light environment.

Light environment may affect insect herbivory through its influence on insect body temperatures (Andrew et al. 2012). As insects are ectotherms, their metabolism and feeding rates are sensitive to small variations in environmental temperature (Frazier et al. 2006). In general, low temperatures reduce the growth, survival and consumption rates of insect herbivores (Scriber and Slansky 1981, Lindroth et al. 1997, Levesque et al. 2002, Paritsis and Veblen 2010). Since welllit leaves are often several degrees warmer than those in the understorey (Pearcy 1987, Naidu and DeLucia 1998), lowlight environments could involve temperature limitations for herbivores in temperate regions; ectotherms do in fact move among different light environments for thermoregulation (Heinrich 1995, Schultz 1998). Given that air temperature decreases in general with latitude, but that the optimal temperature range for insect metabolism is similar across latitudes (Tauber et al. 1987, Frazier et al. 2006, Deutsch et al. 2008, Huey 2010), the search for sunny, warm spots will be more important in temperate than in tropical regions. Notably, while many studies have used latitude to test for temperature effects on global pattern of herbivory (del-Val and Armesto 2010, Garibaldi et al. 2011, Moles et al. 2011, Zhang et al. 2011), no one appears to have considered the influence of light as a moderator of temperature (Fig. 1). An interactive

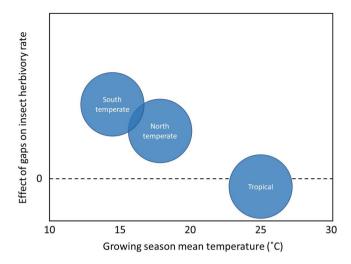


Figure 1. Graphical representation of our first hypothesis, showing predicted effects of growing season mean temperature on the effect of light environment on insect herbivory across different climate types, assuming similar optimum temperatures for insect herbivores across biomes (Frazier et al. 2006, Huey 2010). y-axis shows predicted effect of canopy gaps on herbivory rates in different climate types: insect herbivores are most likely to prefer well-lit environments (gaps) in the oceanic temperate climates of the Southern Hemisphere, where low growing-season temperatures (Lieth et al. 1999) may result in understorey temperatures being well below the optimum for insect metabolism.

effect of light and latitude on herbivory would have important implications for the physiological ecology of forest succession. For example, insect herbivory in the understorey of temperate forests might be insufficient to favour the evolution of herbivory tolerance in shade-tolerant late-successional species – especially in the Southern Hemisphere, where oceanic climates result in lower average growing-season temperatures than in the continental climates of the Northern Hemisphere (Lieth et al. 1999). In contrast, as temperatures in tropical climates are close to the optimum for insect performance (Deutsch et al. 2008), insect herbivory levels may be high enough to select for herbivory tolerance (Kitajima 1994, Myers and Kitajima 2007). In the tropics, herbivores might even prefer the understorey because gaps might expose them to excessive radiative heating (Fig. 1).

Light environment might affect herbivory through its influence on resource availability. Primary productivity in gaps is higher than in understories because plant growth is strongly light-limited in understories (Augspurger 1984, King 1994, Richards and Coley 2007). In some cases, higher light availability also increases leaf nutrient concentrations (e.g. nitrogen) (Osier and Jennings 2007) (but see Lusk and Reich 2000). Well-lit environments may thus attract more insect herbivores than understoreys because of much more abundant and possibly more nutritious foliage (Richards and Coley 2008); this might lead to higher herbivory rates (Richards and Coley 2007). Gaps have especially pronounced effects on the biomass of light-demanding, fast-growing

plants, which respond more strongly to light than shade-tolerant, slow-growing species (Kitajima 1994). On the other hand, chemical and physical defences have been found to increase with light availability (Niesenbaum 1992, Chacón and Armesto 2006, Niesenbaum and Kluger 2006, Salgado-Luarte and Gianoli 2010, Sinimbu et al. 2012), which could lead to the opposite pattern: higher herbivory rates in low-light environments.

In this paper we assess global patterns in the influence of light availability on the incidence of insect damage to leaves, and discuss the implications for plant carbon storage. We gathered studies quantifying insect herbivory within plant species (i.e. intra-specific level) in contrasting light environments, and then conducted a meta-analysis to explore patterns within and across major climatic regions. We considered three hypotheses about controls on global and regional patterns in herbivory. First, that temperature limitation is the primary control on patterns of insect herbivory; in this case the relative importance of understorey herbivory should decrease with latitude, with insect herbivory being higher in gaps than in understories in temperate forests – especially in the oceanic climates of the Southern Hemisphere (Fig. 1). We saw less reason to expect such differences between gap and understorey environments in tropical forests, although we considered that insect herbivores might be stressed by radiative heating in high-light environments (Deutsch et al. 2008) (Fig. 1). Second, we examined the hypothesis that herbivore activity is primarily a function of resource availability (Richards and Coley 2007); in this case, herbivory rates should be higher in gaps than in understoreys regardless of latitude and climate, because of the greater productivity of well-lit environments. If present, such a pattern should be more pronounced in light-demanding species than in shadetolerant associates. Finally, leaf palatability could be the primary control on patterns of insect herbivory; in this case herbivory rates should be lower in gaps than in understoreys, regardless of climate and latitude.

Methodological approach

Literature searches

We conducted a first search on the Web of Science (accessed 25 April 2014) and a second one at 24 April 2016 looking for any type of study in English or Spanish that analysed insect herbivory of plants in sun and shade environments. Specifically, we searched abstracts using the following combination of words: 'herbivory* AND light* AND forest*'. We found 425 studies from which we discarded those using artificial shade, simulated herbivory, non-insect herbivory (e.g. deer browsing), and aquatic herbivory. To be included in our meta-analysis a study should 1) quantify insect herbivory of a terrestrial plant (a single species or group of species) growing naturally under at least two contrasting light environments within a range of light availability. In general, we refer to a

'sun' (i.e. high light) environment as one with an open canopy, and a 'shade' (i.e. low light) environment as one with a closed canopy. Specifically, 'sun' environments included gaps with >10% canopy openness to completely open conditions; 'shade' environments included small gaps (<10% canopy openness) to completely closed forest understorey. When more than one 'shade' environment was considered in a study, we used the data corresponding with the lowest light availability. When different environments were assessed by a study, we avoided data from non-forest ecosystems (e.g. pasture), and opted instead for data from gaps or forest edges. Possible source of bias could arise from differences in the light environments used by different studies. A comparison of the canopy openness (i.e. the most common measure of light environment reported) quoted as 'shade' or as 'sun' in the different studies showed no significant difference amongst the three climates examined (results from one-way ANOVA for shade: $F_{2.18} = 0.66$, p = 0.53; sun: $F_{2.18} = 0.20$, p = 0.82; n = 4; SH temperate, n=5; NH temperate, n=10 tropical). This suggests that the light environments were overall comparable amongst climates. Most studies meeting our criteria used tree seedlings, saplings, and adults, whereas a limited number of the studies included used herbs or shrubs. We found some species (e.g. Lindera benzoin) and study sites (e.g. Barro Colorado) to be overrepresented relative to others (Table 1). When a study evaluated herbivory in more than one species, we included them as separate cases (i.e. replicates). When herbivory was measured on more than one date, we averaged across dates. Finally, when different treatments were applied in the different light environments (e.g. addition of fertilizer, herbivore exclusion) we only used the data from the controls. Our final list included 40 published studies and one unpublished study, covering 87 cases (i.e. comparisons) of intra-specific insect herbivory between contrasting light environments; 40 studies covered 70 plant species (a same species was sometimes used in two or more studies, Table 1) and one study covered six pooled species (Goodale et al. 2014); 38 studies used woody species (69 species) and three studies used herbaceous species (three species) (Liang and Stehlik 2009, Agrawal et al. 2012, Jones and Klemetti 2012) (Table 1). Geographically, the studies covered the temperate Northern Hemisphere (34° to 57°N, 32 cases), the temperate Southern Hemisphere (40° to 46°S, 12 cases), the subtropics (31°N to 33°S, one case which was treated as temperate) and the tropics (Northern and Southern Hemisphere) (20°N to 08°S, 42 cases) (Table 1). There were 23 cases of shadeintolerant species, 27 cases of semi-tolerant species, and 37 cases of shade-tolerant species (Table 1).

We extracted or estimated the following information from each selected study: name of plant species; level of herbivory for each light environment; light availability for each light environment; latitude; climate type (i.e. temperate, tropical); mean temperature for the first summer month (July for the Northern Hemisphere; January for the Southern Hemisphere); and ontogenetic stage of studied plants. Our study list included diverse measurements of herbivory

Table 1. List of studies and species (i.e. cases) included in our meta-analysis. In all cases insect herbivory was evaluated in two contrasting light environments (shade, canopy openness < 10%; sun, canopy openness > 10%). For each case, the source of data extraction for meta-analysis (i.e. text, table or figure), the latitude, the climate type, the mean temperature of the warmest month (estimated by DIVA-GIS, ver. 7.5.0.0), the shade tolerance (i: intolerant; m: semitolerant; t: tolerant), the leaf habit (D: deciduous; E: evergreen, H: herb), ontogeny, and light environments considered, are indicated.

| Study | Source | Latitude | Climate | Summer T (°C) | Species | Tolerance | Habit | Ontogeny | Shade criteria | Sun criteria |
|------------------------------------|---------------------|------------|----------|------------------|-----------------------------------|-----------|---------|----------|---|--|
| Prather (2014) | Fig. 3, 4 | 18°19′00″N | tropical | 22.5 | Miconia prasina | | E tree | sapling | < 10% CO | 20-30% CO |
| Prather (2014) | Fig. 3, 4 | 18°19′00″N | tropical | 22.5 | Miconia racemosa | | E tree | sapling | < 10% CO | 20-30% CO |
| Prather (2014) | Fig. 3, 4 | 18°19′00″N | tropical | 22.5 | Schefflera morototoni | | E tree | sapling | < 10% CO | 20-30% CO |
| Prather (2014) | Fig. 3, 4 | 18°19′00″N | tropical | 22.5 | Casearia arborea | t | E tree | sapling | < 10% CO | 20-30% CO |
| Prather (2014) | Fig. 3, 4 | 18°19′00″N | tropical | 22.5 | Sloanea berteriana | t | E tree | sapling | < 10% CO | 20-30% CO |
| Prather (2014) | Fig. 3, 4 | 18°19′00″N | tropical | 22.5 | Manilkara bidentata | t | E tree | sapling | < 10% CO | 20-30% CO |
| Angulo-Sandoval and Aide (2000) | Fig. 5 | 18°20′00″N | tropical | 22.5 | Manilkara bidentata | t | E tree | seedling | 81.4 µmol m ⁻² s ⁻¹ | 737.1 µmol m ⁻² s ⁻¹ |
| Lopez-Toledo et al. (2008) | Fig. 1d | 16°07′12″N | tropical | 27.2 | Brosimum alicastrum | t | E tree | sapling | <1.5 crown index | >2.5 crown index ¹⁴ |
| Gerhardt (1998) | Fig. 3 ² | 10°51′00″N | tropical | 25.0 | Swietenia macrophylla | Е | D tree | seedling | 5-10% PAR | 35-45% PAR |
| Gerhardt (1998) | Fig. 3 ² | 10°51′00″N | tropical | 25.0 | Cedrela odorata | | D tree | seedling | 5-10% PAR | 35-45% PAR |
| Letourneau and Dyer (1998) | Fig. 3 | 10°26′00″N | tropical | 26.1 | Piper cenocladum | t | E shrub | sapling | 91.7% CC | 83.3% CC |
| Dyer and Letourneau (1999) | Fig. 2b | 10°26′00″N | tropical | 26.1 | Piper cenocladum | t | E shrub | sapling | >96.6% cover | <91.7% cover |
| de la Fuente and Marquis (1999) | Fig. 2 | 09°05′00″N | tropical | 26.5 | Stryphnodendron microstachyum | t | E tree | sapling | secondary forest | open pasture |
| Harrison (1987) | text | 09°10′00″N | tropical | 26.5 | Hybanthus prunifolius | Е | E shrub | sapling | understorey | gaps |
| Coley (1983) | Table 1 | 09°10′00″N | tropical | 26.5 | Trichilia cipo | Ш | E tree | sapling | understorey | 100-800 m ² gaps |
| Pearson et al. (2003a) | Fig. 4 ³ | 09°10′00″N | tropical | 26.5 | Miconia argentea | Е | D tree | seedling | ₉ OO %9 | 15% CO ¹⁹ |
| Pearson et al. (2003a) | Fig. 4 ³ | 09°10′00″N | tropical | 26.5 | Cecropia insignis | Е | E tree | seedling | ₉ OO %9 | 15% CO ¹⁹ |
| Pearson et al. (2003a) | Fig. 4 ³ | 09°10′00″N | tropical | 26.5 | Trema micrantha | | E tree | seedling | 9OO %9 | 15% CO ¹⁹ |
| Richards and Coley (2008) | Table 2 | 09°05′00″N | tropical | 26.5 | Hybanthus prunifolius | Ε | E shrub | sapling | understorey | gap |
| Pearson et al. (2003b) | Fig. 3b | 09°05′00″N | tropical | 26.5 | Miconia argentea | t | D tree | seedling | 3.32% PAR | 26.4% PAR |
| Pearson et al. (2003b) | Fig. 3b | 09°05′00″N | tropical | 26.5 | Cecropia insignis | Е | E tree | seedling | 3.32% PAR | 26.4% PAR |
| Pearson et al. (2003b) | Fig. 3b | 09°05′00″N | tropical | 26.5 | Luehea seemannii | | D tree | seedling | 3.32% PAR | 26.4% PAR |
| Pearson et al. (2003b) | Fig. 3b | 09°05′00″N | tropical | 26.5 | Trema micrantha | | E tree | seedling | 3.32% PAR | 26.4% PAR |
| Pearson et al. (2003b) | Fig. 3b | 09°05′00″N | tropical | 26.5 | Ochroma pyramidale | | E tree | seedling | 3.32% PAR | 26.4% PAR |
| Pearson et al. (2003b) | Fig. 3b | 09°05′00″N | tropical | 26.5 | Croton bilbergianus | t | E tree | seedling | 3.32% PAR | 26.4% PAR |
| Goodale et al. (2014) | Table 2 | 06°23′23″N | tropical | 22.9 | Dillenia triquetra and five other | Ε | E tree | seedling | 0.08 visual sky ¹⁰ | 0.36 visual sky ¹⁰ |
| Norghauer and Newbery (2013) | Fig. 34 | 05°17′09″N | tropical | 25.0 | Microberlinia bisulcata | | D tree | seedling | 5.53% CO ¹¹ | 7.69% CO ¹⁶ |
| Norghauer and Newbery (2013) | Fig. 34 | 05°17′09″N | tropical | 25.0 | Tetraberlinia bifoliolata | + | E tree | seedling | 5.48% CO ¹² | 8.23% CO ¹⁷ |

| Norghauer and Newbery (2013) Eichhorn et al. (2010) | | Latitude | Climate | Summer T (°C) | Species | Tolerance | Habit | Ontogeny | Shade criteria | Sun criteria |
|---|--------------------|------------|-------------|------------------|----------------------------|-----------|--------|----------|--------------------------|------------------------|
| Eichhorn et al. (2010) | Fig 3 ⁴ | 05°17′09″N | tropical | 25.0 | Tetraberlinia korupensis | Е | E tree | seedling | 5.64% CO ¹³ | 9.21% CO ¹⁸ |
| | Fig. 1c, d | 05°10′N | tropical | 26.8 | Hopea nervosa | + | D tree | seedling | understorey | 400–600 m² gap |
| Eichhorn et al. (2010) | Fig. 1c, d | 05°10′N | tropical | 26.8 | Parashorea tomentella | ţ | D tree | seedling | understorey | 400-600 m² gap |
| Eichhorn et al. (2010) | Fig. 1c, d | 05°10′N | tropical | 26.8 | Shorea fallax | + | E tree | seedling | understorey | 400-600 m² gap |
| Eichhorn et al. (2010) | Fig. 1c, d | 05°10′N | tropical | 26.8 | Shorea multiflora | ţ | E tree | seedling | understorey | 400–600 m² gap |
| Eichhorn et al. (2010) | Fig. 1c, d | 05°10′N | tropical | 26.8 | Hopea beccariana | ţ | E tree | seedling | understorey | 400–600 m² gap |
| Whitmore and Brown (1996) | Fig. 3b | 04°54′00″N | tropical | 24.8 | Hopea nervosa | ţ | D tree | seedling | 6% CO ⁷ | 26% CO ²² |
| Whitmore and Brown (1996) | Fig. 3b | 04°54′00″N | tropical | 24.8 | Parashorea malaanonan | | E tree | seedling | 6% CO ⁷ | 26% CO ²² |
| Whitmore and Brown (1996) | Fig. 3b | 04°54′00″N | tropical | 24.8 | Shorea johorensis | | E tree | seedling | 6% CO ⁷ | 26% CO ²² |
| Norghauer et al. (2008) | Fig. 3b | 07°46′14″S | tropical | 25.2 | Swietenia macrophylla | ш | D tree | seedling | 7.1% CO ⁸ | 14.1% CO ²³ |
| Benitez-Malvido et al. (2005) | Table 1 | 03°06′00″S | tropical | 27.4 | Chrysophyllum pomiferum | + | E tree | seedling | 3.8% PAR | 15.5% PAR |
| Benitez-Malvido et al. (2005) | Table 1 | 03°06′00″S | tropical | 27.4 | Micropholis venulosa | + | E tree | seedling | 3.8% PAR | 15.5% PAR |
| Benitez-Malvido et al. (2005) | Table 1 | 03°06′00″S | tropical | 27.4 | Pouteria caimito | + | E tree | seedling | 3.8% PAR | 75.6% PAR |
| Sinimbu et al. (2012) | Fig. 1h | 02°24′00″S | tropical | 27.0 | Inga paraensis | | E tree | sapling | <10% CO | >10% CO |
| Christie and Hochuli (2005) | Fig. 2 | 33°52′06″S | subtropical | 21.5 | Angophora costata | | E tree | adult | interior | edge |
| Löf et al. (2005) | Fig. 5 | 57°05′N | temp N | 14.6 | Fagus sylvatica | Ļ | D tree | seedling | 5% full light | 20% full light |
| Löf et al. (2005) | Fig. 5 | 57°05′N | temp N | 14.6 | Quercus robur | Е | D tree | seedling | 5% full light | 20% full light |
| Löf et al. (2005) | Fig. 5 | 57°05′N | temp N | 14.6 | Tilia cordata | E | D tree | seedling | 5% full light | 20% full light |
| Löf et al. (2005) | Fig. 5 | 57°05′N | temp N | 14.6 | Acer platanoides | t | D tree | seedling | 5% full light | 20% full light |
| Löf et al. (2005) | Fig. 5 | 57°05′N | temp N | 14.6 | Fraxinus excelsior | Е | D tree | seedling | 5% full light | 20% full light |
| Löf et al. (2005) | Fig. 5 | 57°05′N | temp N | 14.6 | Prunus avium | | D tree | seedling | 5% full light | 20% full light |
| Löf et al. (2005) | Fig. 5 | 57°05′N | temp N | 14.6 | Picea abies | ţ | E tree | seedling | 5% full light | 20% full light |
| Karolewski et al. (2013) | Fig. 2 | 52°33′00″N | temp N | 16.5 | Sambucus nigra | Ε | D tree | adult | $understorey^9$ | open ²¹ |
| Karolewski et al. (2013) | Fig. 2 | 52°33′00″N | temp N | 16.5 | Cornus sanguinea | Е | D tree | adult | understorey ⁹ | open ²¹ |
| Karolewski et al. (2013) | Fig. 2 | 52°33′00″N | temp N | 16.5 | Frangula alnus | ш | D tree | adult | understorey ⁹ | open ²¹ |
| Karolewski et al. (2013) | Fig. 2 | 52°33′00″N | temp N | 16.5 | Corylus avellana | t t | D tree | adult | understorey ⁹ | open ²¹ |

| Study | Source | Latitude | Climate | Summer T (°C) | Species | Tolerance | Habit | Ontogeny | Shade criteria | Sun criteria |
|--|---------------------|--------------------------|----------------------------|------------------|--------------------------------------|------------|--------------------|----------------|---|---|
| Karolewski et al. | Fig. 2 | 52°33′00″N | temp N | 16.5 | Prunus serotina | | D tree | adult | understorey ⁹ | open ²¹ |
| (2013) Karolewski et al. (2013) | Fig. 2 | 52°33′00″N | temp N | 16.5 | Prunus padus | E | D tree | adult | $understorey^9$ | open ²¹ |
| McGeoch and Gaston | Table 3 | 53°20′31″N | temp N | 14.2 | llex aquifolium | ٤ | E tree | adult | interior | edge |
| Liang and Stehlik (2009) | Fig. 2 | 44°01′46″N | temperate N | 24.4 | Asarum canadense | ÷ | エ | adult | >91% CC | <85% CC |
| Agrawal et al. (2012) Niesenbaum and Kluger (2006) | Fig. 1d Fig. 2A | 42°27′27″N 40°40′13″N | temperate N temperate N | 20.5 | Asclepias syriaca Lindera benzoin | t B | H D shrub | adult adult | 168 µmol m ⁻² s ⁻¹ 14.39 µmol m ⁻² s ⁻¹ | 1784 µmol m ⁻² s ⁻¹ 214.08 µmol m ⁻² s ⁻¹ |
| Niesenbaum (1992) Muth et al. (2008) | Table 2 Fig. 1 | 40°40′13″N 40°40′13″N | temperate N temperate N | 19.5 | Lindera benzoin Lindera benzoin | + + | D shrub D shrub | adult adult | 0.86% full light ¹⁵ 48 µmol m ⁻² s ⁻¹ | 19.84% full light ²⁴ 658.30 µmol m ⁻² |
| Lieurance and | Fig. 1a, c | 40°1-39°3N | temperate N | 20.7 | Lonicera maackii | Е | D shrub | adult | interior | edge |
| Jones and Klemetti | Fig. 2 | 41°57′59″N | temperate N | 14.4 | Trollius laxus | | エ | adult | $<150\ \mu mol.m^{-2}.s^{-1}$ | $> 250 \mu mol m^{-2}$ |
| Arrieta and Suárez | Fig. 4 ⁵ | 41°57′59″N | temperate N | 14.4 | llex aquifolium | Е | E tree | seedling | 0.70% PPFD | 46.59% PPFD |
| (2003) Stoepler and Rehill (2012) | Fig. 36 | 39°15′9′N | temperate N | 21.1 | Quercus alba | Е | D tree | sapling | 2.4% CO | 57.9% CO |
| Lincoln and Mooney (1984) | Fig. 1 | 37°24′N | temperate N | 20.0 | Diplacus aurantiacus | ш | D shrub | adult | <15 mol m ⁻² d ⁻¹ | >40 mol m ⁻² d ⁻¹ |
| Shure and Wilson (1993) | Fig. 6 | 35°12′43″N | temperate N | 19.3 | Robinia pseudoacacia | | D tree | sapling | understorey | 0.08 ha's gap |
| Shure and Wilson (1993) | Fig. 6 | 35°12′43″N | temperate N | 19.3 | Cornus florida | + | D tree | sapling | understorey | 0.08 ha's gap |
| Shure and Wilson (1993) | Fig. 6 | 35°12′43″N | temperate N | 19.3 | Liriodendron tulipifera | | D tree | sapling | understorey | 0.08 ha's gap |
| Shure and Wilson (1993) | Fig. 6 | 35°12′43″N | temperate N | 19.3 | Acer rubrum | ţ | D tree | sapling | understorey | 0.08 ha's gap |
| Hunter and Forkner (1999) | Fig. 5a | 35°N | temperate N | 19.3 | Acer rubrum | t t | D tree | sapling | interior | 0.2-0.4 ha open area |
| Hunter and Forkner (1999) | Fig. 5b | 35°N | temperate N | 19.3 | Quercus rubra | | D tree | sapling | interior | 0.2-0.4 ha open area |
| Dudt and Shure (1994) | Fig 6. | 34°41′37″N | temperate N | 22.1 | Liriodendron tulipifera | | D tree | seedling | 6% full light | $80 \text{ m}^2 \text{ gap}^{25}$ |
| Dudt and Shure (1994) | | 34°41′37″N | temperate N | 22.1 | Cornus florida | t | D tree | seedling | 6% full light | $80 \text{ m}^2 \text{ gap}^{25}$ |
| Piper, F. unpubl. | | 46°29′51″S | temperate S | 12.9 | Amomyrtus luma | t | E tree | seedling | < 7 % CO | > 10% CO |
| Piper, F. unpubl. | | 46°29′51″S | temperate S | 12.9 | Drymis winteri | ш | E tree | seedling | < 7 % CO | > 10% CO |

Table 1. Continued

Table 1. Continued

| | | | | Summer | | | | | | |
|--------------------------------------|---------|------------|-------------|--------|-----------------------|-----------|---------|--------------------------|----------------|----------------|
| Study | Source | Latitude | Climate | T (°C) | Species | Tolerance | Habit | Tolerance Habit Ontogeny | Shade criteria | Sun criteria |
| Piper, F. unpubl. | | 45°27′07″S | temperate S | 11.7 | Aristotelia chilensis | | E tree | seedling | < 7 % CO | > 10% CO |
| Piper, F. unpubl. | | 45°27′07″S | temperate S | 11.7 | Amomyrtus luma | ţ | E tree | seedling | < 7 % CO | > 10% CO |
| Piper, F. unpubl. | | 45°27′07″S | temperate S | 11.7 | Azara lanceolata | Ε | E tree | seedling | < 7 % CO | > 10% CO |
| Piper, F. unpubl. | | 45°27′07″S | temperate S | 11.7 | Luma apiculata | t | E tree | seedling | < 7 % CO | > 10% CO |
| Bach and Kelly (2004) text | text | 43°09′S | temperate S | 11.0 | Alepis flavida | t | E shrub | adult | interior | edge |
| Chacón and Armesto (2006) | Fig. 3 | 41°53′00″S | temperate S | 12.4 | Drimys winteri | E | E tree | seedling | 1% CO | 11.5% CO |
| Chacón and Armesto (2006) | Fig. 3 | 41°53′00″S | temperate S | 12.4 | Gevuina avellana | Ε | E tree | seedling | 1% CO | 11.5% CO |
| Piper, F. unpubl. | | 41°35′00″S | temperate S | 15.2 | Amomyrtus meli | ţ | E tree | sapling | < 7 % CO | > 10% CO |
| Salgado-Luarte and Gianoli (2010) | Fig. 1A | 40°39′00″S | temperate S | 15.2 | Embothrium coccineum | | E tree | seedling | 5% full light | 67% full light |
| Salgado-Luarte and Gianoli (2012) | Fig. 1A | 40°39′00″S | temperate S | 15.2 | Aristotelia chilensis | | E tree | seedling | 5.1% CO | 25.6% CO |

¹ mean temperature of July for the Northern Hemisphere and January for Southern Hemisphere ² average for two forests (deciduous and semievergreen); Th0Tr0 only

outside exclosures only

4,6 controls only

8 4.5 mol m⁻² day⁻¹

⁵ 1997s cohort; edge versus closed holly woodland ⁶ 0.2 mol m² day⁻¹, 2.83% PAR, 0.13 GSF ⁷ 1 mol m⁻² day⁻³

¹⁰ proportion at 180°
¹¹ 0.59% PPF
¹² 0.73% PPFD
¹³ 1.10% PPFD
¹⁴ gaps >200 m²
¹⁵ 9.18 μmol m⁻² s⁻¹

⁹ under a canopy of Pinus sylvestris with admixture of Quercus robur, Fagus sylvatica, Carpinus betulus and Ulmus laevis

¹⁶ 5.36% PPFD ¹⁷ 5.39% PPFD ¹⁸ 6.04% PPFD ¹⁹ 8 mol m⁻² day⁻¹, 26.9% PAR, 0.27 GSF

 21 along wide forest roads or in forest gaps 22 19 mol m 22 day $^{-1}$ 23 12.8 mol m $^{-2}$ day $^{-1}$

²⁴ 115.68 mmol m⁻² s⁻¹

 25 24% full light For Prather (2014) "shade" was "trim, no debris", and "sun" was "no trim, no debris"

(e.g. herbivory index, % leaf area removed), mostly corresponding to folivory (Supplementary material Appendix 1 Table A1). The mean values for herbivory, standard deviations, confidence intervals or standard errors (for standard deviation calculation), and sample size were extracted for each light environment from the text, the tables or the figures, in the latter case using the software TechDig (ver. 2.0, Ronald B. Jones). When the results did not include the standard deviations, confidence intervals or standard errors, we attempted to contact the authors for the information; two studies had to be omitted because of no response. When the sampling unit was not clearly stated, we used the largest scale sampling unit (i.e. sites over plots; plots over plants; plants over leaves) and in some cases averaged values from smaller scales to generate larger scale values (e.g. leaves to plant). Here, we assumed independence among sample units. Some studies considered different individuals of a same plot, or different leaves of same plants, as replicates. In these cases, we averaged herbivory values of plants of a same plot or leaves of a same plant and considered it as a replicate. Latitude was recorded using the geographic coordinates reported in the study or Google Earth (ver. 7.1.5.1557) when they were not provided. The mean temperature for the first summer month was estimated for each study site using Worldclim (ver. 1, 30 sec ESRI data bases) (Hijmans et al. 2005) and DIVA-GIS software (ver. 7.5.0.0). We used the first summer month (i.e. the warmest) for our temperature measurement because it is generally the period of seedling establishment, the maximum expansion rate of new leaves, and the maximum rate of herbivory in temperate forests (Coley and Aide 1991, Lowman 1992, Carus 2009). More importantly, this is usually the month for which herbivory data were reported in most studies.

Leaf habit (deciduous or evergreen) and shade tolerance (shade-tolerant, semi-tolerant, shade-intolerant) of the studied plant species were obtained from the original publications and from web databases. The latter included The Plant List (< www.theplantlist.org/>), Encyclopedia of Life (< http://eol.org/>), Global Species (< www.globalspecies. org/>), Integrated Taxonomic Information System (< www.tiss.gov/>), and Global Biodiversity Information Facility (< www.gbif.org/>).

Data analyses

For each case, we quantified the difference in herbivory damage between shade and sun environments as the effect size. An effect size is a standardized measure of the magnitude of the observed effect. Thus, effect sizes across studies that have measured herbivory in diverse ways and in diverse units or scales, can be directly analysed (Field and Gillett 2010). We chose the standardized mean difference, specifically 'Hedges's g', as a metric of the effect size, defined as the difference between the means of two groups divided by their pooled standard deviation (Gurevitch and Hedges 1993). This number is then multiplied by a correction factor J, to correct for small-sample bias. In our case, 'sun' herbivory was considered

as the 'experimental group' and 'shade' herbivory as the 'control group'. Thus, a positive effect size means that herbivory is higher in sun than in shade, while a negative effect size means the opposite. The effect size was calculated as:

$$dij = \left(X^{sun}_{ij} - X^{shade}_{ij}\right) / S_{ij} \times J$$

where d is calculated for the jth study in the ith study class; X^{sun}_{ij} is the mean of the sun group; X^{shade}_{ij} is the mean of the shade group; S_{ij} is the pooled standard deviation of the sun and shade groups; and J is a correction factor for small sample sizes. Our analyses did not correct a priori for publication bias because most of the selected studies did not address the difference in herbivory between light environments as a target research question. More often they treated the effect of light environment on the growth or abundance of plants, the performance of insect herbivores, or the interaction of light environment with a particular treatment (e.g. fertilization) on the performance or abundance of herbivores.

We first tested the effect of light environment on insect herbivory on a global scale (i.e. across climates), by comparing the effect sizes between 'sun' and 'shade', using fully random-effects analyses. Then, climate type and shade tolerance were treated as moderators and their effects were examined using mixed effects analysis. Climate was tested based on the following sub-groups: northern temperate, southern temperate and tropical (the only subtropical study was treated as SH temperate). Shade tolerance was tested based on the following sub-groups: shade-tolerant, shade-intolerant, shade semi-tolerant. A random effects model was used to combine studies within each subgroup (i.e. climate or shade tolerance category), and a fixed effect model was used to combine subgroups and yield the overall effect. The study-to-study variance (tau-squared) was assumed to be the same for all subgroups - this value was computed within subgroups and then pooled across subgroups. Mixed-effects models are more appropriate than fixed-effects models for making inferences that generalize beyond the studies included in the metaanalysis (Field and Gillett 2010). They assume that the average effect size in a population varies randomly from study to study: studies in a meta-analysis are made up of populations with different average effect sizes (Field and Gillett 2010). Mixed-effect models consider two sources of error: 1) error created by sampling studies from a population, and 2) error created by sampling individual populations from a universe of populations (Field and Gillett 2010).

Latitude and temperature were also considered as moderators and their effects evaluated by meta-regression. Latitude was examined either distinguishing or not between hemispheres (i.e. using negative sign for the Southern Hemisphere and positive sign for the norther hemisphere, or using always positive values, respectively). Meta-regression under randomeffect model was also used to assess the combined impact of climate, latitude, temperature and shade tolerance. All analyses were performed with comprehensive meta-analysis software (ver. 3.3.070, 2014).

Results

Globally, there were 19 cases of higher insect herbivory in the sun, 10 cases of higher herbivory in the shade, and 58 cases of no difference between contrasting light environments (Supplementary material Appendix 1 Table A2). The mean effect size of the complete data set was positive and just significantly different from zero (d=0.221, SE=0.113, 95% CI = 0.000 - 0.443, n = 87; Z-value = 1.957, p-value = 0.050), indicating that insect herbivory at a global scale was marginally but significantly higher in sun than in shade (Fig. 2). There was no significant overall effect of climate type on the herbivory at sun versus shade (Q-value=4.132, df=2, p = 0.127). However, when climates were analysed separately (i.e. as subgroups), herbivory in the tropics was significantly higher in the sun than in the shade (i.e. positive effect size) (d=0.413, SE=0.162; 95% CI=0.026-0.097, n=42;Z=2.558, p=0.011) (Fig. 2). In both northern and southern temperate zones, insect herbivory was similar between

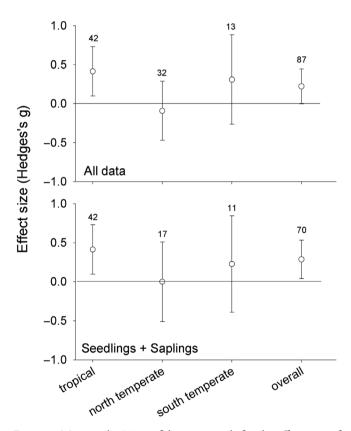


Figure 2. Mean and 95% confidence intervals for the effect sizes of light environment (sun versus shade, Table 1) on insect herbivory (weighted standardized mean, Hedges's g) for all study cases (i.e. a study \times species' combination; $n\!=\!87$), study cases of tropical forests ($n\!=\!42$), study cases of north temperate forests ($n\!=\!32$), and study cases of south temperate forests ($n\!=\!13$). Positive values indicate preference for sunny over shady environments. Upper panel shows results for the complete data set, including seedlings, saplings and adults; lower panel shows results for seedlings and saplings (i.e. subgroup 'adults' excluded).

light environments (Supplementary material Appendix 1 Table A2, Fig. 2).

Most studies were conducted on seedlings or saplings, with a lower proportion of studies conducted on adult plants. The latter, however, represented ca 50% of the case studies in Northern Hemisphere temperate forests, but 0% of the tropical cases. To control for possible ontogenetic effects on our results, we re-ran our analyses excluding adult plants. The mean effect size in this case was positive and significantly different from zero (d=0.287, SE=0.123, 95% CI=0.045-0.529, n=70; test of the null: Z=2.326, p=0.020). Also, climate type had no significant overall effect on herbivory in sun versus shade (Q=1.860, df=2, p=0.395), but herbivory was higher in sun than in shade in the tropics (d=0.413, SE=0.162; 95% CI=0.096–0.731, n=42; Z=2.552, p=0.011) and similar between light environments for the other biomes (Fig. 2).

Globally, species shade tolerance had no effect on the relative levels of herbivory in sun and shade environments (Q=1.405, df=2, p=0.495). However, as a group, shade-intolerant species suffered more herbivory in the sun than in the shade (d=0.428, SE=0.216, 95% CI=0.004-0.852, n=23; Z=1.979, p=0.049), while herbivory rates suffered by semi-tolerant and shade-tolerant species did not differ between light environments (Fig. 3).

Variation in effect sizes was not explained by temperature or latitude (Table 2). Models including different combinations of moderators also failed to explain the variation in effect sizes (Table 2). However, 'climate' had a marginally significant effect in model 8. Temperature and latitude, and different combinations of moderators, also failed to explain the variation in effect sizes for studies considering seedlings and saplings only (Table 3). The moderator 'climate' had a significant effect in two models (model 5 and 8, Table 3).

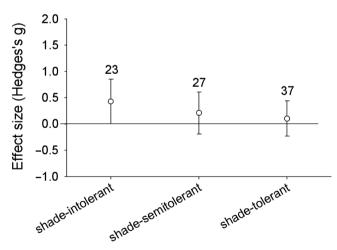


Figure 3. Mean and 95% confidence intervals for the effect sizes of light environment (sun versus shade, Table 1) on insect herbivory (weighted standardized mean, Hedges's g) for case studies conducted in shade-intolerant species (n=23), shade semi-tolerant (n=27) and shade-tolerant (n=37). Negative values indicate preference for sunny over shady environments.

Table 2. Effects of different moderators on the effect sizes of insect herbivory in sun versus shade for all case studies (Table 1), examined by meta-regression. Tau² (variance of true effect sizes) and l² (between-study variance, i.e. that can be potentially explained by additional study-level covariates) indicate the variance that is not explained by the model (goodness of fit).

| Model | Moderators | Q-value | df | р | R^2 | Tau ² | I^2 | Details |
|-------|--|---------|----|-------|-------|------------------|-------|--|
| 1 | Temperature | 0.72 | 1 | 0.396 | 0.00 | 0.86 | 86% | |
| 2 | Climate type | 4.13 | 2 | 0.127 | 0.00 | 0.86 | 85% | |
| 3 | Latitude 1 | 1.45 | 1 | 0.228 | 0.00 | 0.86 | 85% | |
| 4 | Latitude 2 | 2.41 | 1 | 0.121 | 0.03 | 0.81 | 85% | |
| 5 | Temperature Climate type | 4.23 | 2 | 0.121 | 0.02 | 0.83 | 85% | Coef. = -0.05 (0.04), p = 0.248 Coef. = 0.81 (0.43), p = 0.060 |
| 6 | Temperature Latitude 2 | 4.30 | 2 | 0.117 | 0.12 | 0.74 | 83% | Coef. = -0.06 (0.05), p = 0.185 Coef. = -0.25 (0.01), p = 0.060 |
| 7 | Temperature Climate type Latitude 2 | 4.84 | 3 | 0.184 | 0.11 | 0.74 | 84% | Coef. = -0.06 (0.05), p = 0.181 Coef. = 0.50 (0.65), p = 0.445 Coef. = -0.01 (0.02), p = 0.536 |
| 8 | Temperature Climate type Shade tol. | 6.10 | 4 | 0.182 | 0.06 | 0.78 | 84% | Coef. = -0.05 (0.04), p=0.237 Coef. = 0.83 (0.43), p=0.051 Q= 1.76, df=2, p=0.415 |
| 9 | Temperature Climate type Shade tol Latitude 2 | 6.52 | 5 | 0.259 | 0.10 | 0.75 | 83% | Coef. = -0.06 (0.05); p=0.204 Coef. = 0.58 (0.66); p=0.38 Q=1.70, df=2, p=0.43 Coef. = -0.01 (0.02), p=0.63 |

Latitude 1 was computed always positive, i.e. not distinguishing between hemispheres; Latitude 2 included positive and negative values for Northern and Southern Hemispheres, respectively. Model 2 considered three subgroups in *Climate type*, ('tropical', 'NH temperate', 'SH temperate'); models 5, 7, 8 and 9 considered two soubgroups ('temperate' and 'tropical').

When models were run separately for each climate type, we found a negative effect of temperature in tropical climates, indicating that the effect of gaps on herbivory decreases with increasing temperature (Table 4). In temperate forests, however, neither temperature nor latitude explained the variation in effect size.

Discussion

Our meta-analysis did not support our first hypothesis, that temperature limitation is the primary control on patterns of insect herbivory. Assuming that in temperate regions temperatures are suboptimal for most insects (Frazier et al. 2006,

Table 3. Effects of different moderators on the effect sizes of insect herbivory in sun versus shade for case studies including seedlings and saplings only (Table 1), examined by meta-regression (n = 70, Table 1). Tau² (variance of true effect sizes) and l² (between-study variance, i.e. that can be potentially explained by additional study-level covariates) indicate the variance that is not explained by the model (goodness of fit).

| Model | Moderators | Q-value | df | р | R^2 | Tau ² | l^2 | Details |
|-------|--|---------|----|-------|-------|------------------|-------|--|
| 1 | Temperature | 0.14 | 1 | 0.710 | 0.00 | 0.85 | 87% | |
| 2 | Climate type | 1.86 | 2 | 0.395 | 0.00 | 0.86 | 86% | |
| 3 | Latitude 1 | 0.08 | 1 | 0.782 | 0.00 | 0.86 | 85% | |
| 4 | Latitude 2 | 0.77 | 1 | 0.379 | 0.00 | 0.81 | 86% | |
| 5 | Temperature Climate type | 4.56 | 2 | 0.102 | 0.01 | 0.80 | 86% | Coef. = -0.09 (0.05), p=0.086 Coef. = 1.19 (0.56), p=0.035 |
| 6 | Temperature Latitude 2 | 2.92 | 2 | 0.232 | 0.17 | 0.68 | 83% | Coef. = -0.09 (0.06), p=0.153 Coef. = -0.03 (0.02), p=0.096 |
| 7 | Temperature Climate type Latitude 2 | 5.10 | 3 | 0.165 | 0.14 | 0.70 | 84% | Coef. = -0.10 (0.06), p = 0.110 Coef. = 1.05 (0.70), p = 0.133 Coef. = -0.01 (0.02), p = 0.765 |
| 8 | Temperature Climate type Shade tol. | 6.49 | 4 | 0.165 | 0.14 | 0.70 | 84% | Coef. = -0.09 (0.05), p = 0.071 Coef. = 1.23 (0.55), p = 0.026 Q = 1.72, df = 2, p = 0.420 |
| 9 | Temperature Climate type Shade tol Latitude 2 | 6.84 | 5 | 0.232 | 0.14 | 0.70 | 83% | Coef. = -0.09 (0.06); p = 0.128 Coef. = 0.58 (0.66); p = 0.38 Q = 1.78, df = 2, p = 0.41 Coef. = -0.00 (0.02), p = 0.91 |

Latitude 1 was computed always positive, i.e. not distinguishing between hemispheres; Latitude 2 included positive and negative values for Northern and Southern Hemispheres, respectively. Model 2 considered three subgroups in *Climate type*, ('tropical', 'NH temperate', 'SH temperate'); models 5, 7, 8 and 9 considered two soubgroups ('temperate' and 'tropical').

Table 4. Effects of temperature, latitude, and the combination of both on the effect sizes of insect herbivory in sun versus shade in tropical (n=42, Table 1) and temperate forests (n=28, Table 1), examined by meta-regression. Only case studies including seedlings and saplings were analyzed. Tau² (variance of true effect sizes) and I^2 (between-study variance, i.e. that can be potentially explained by additional study-level covariates) indicate the variance that is not explained by the model (goodness of fit).

| Model | Moderators | Q-value | df | р | R^2 | Tau ² | I^2 | Details |
|-----------|-------------------------|---------|----|-------|-------|------------------|-------|--|
| tropical | | | | | | | | |
| 1 | Temperature | 2.31 | 1 | 0.129 | 0.07 | 0.98 | 89% | |
| 2 | Latitude | 0.13 | 1 | 0.718 | 0.02 | 0.99 | 72% | |
| 3 | Temperature Latitude | 4.03 | 2 | 0.133 | 0.30 | 0.75 | 85% | Coef. = -0.25 (0.13), p = 0.048 Coef. = -0.04 (0.04), p = 0.304 |
| temperate | | | | | | | | · |
| 1 | Temperature | 1.34 | 1 | 0.247 | 0.00 | 0.39 | 72% | |
| 2 | Latitude | 0.78 | 1 | 0.377 | 0.00 | 0.39 | 72% | |
| 3 | Temperature Latitude | 1.37 | 2 | 0.503 | 0.00 | 0.40 | 72% | Coef. = 0.06 (0.06), p = 0.279 Coef. = -0.00 (0.00), p = 0.789 |

Deutsch et al. 2008, Huey 2010), and in view of a general positive correlation between light availability and temperature (Andrew et al. 2012), we had predicted that the relative importance of understorey herbivory should decrease with latitude, with insect herbivory being higher in gaps than in understories in temperate forests – especially in the oceanic climates of the Southern Hemisphere (Fig. 1). Contrary to our prediction, our meta-analysis found that insect herbivory was lower in the understorey than in gaps in general, and in tropical forests in particular (Fig. 2). The fact that preferential carbon allocation to storage in late-successional species is more characteristic of tropical than of temperate forests (Kitajima 1994, Canham et al. 1999, Lusk and Piper 2007, Myers and Kitajima 2007, Poorter and Kitajima 2007, Piper 2015) is therefore unlikely to be explained by differential herbivore responses to light availability gradients in tropical versus temperate forests. Although the low opportunity cost of allocation to storage in shaded understories has been emphasized (Kobe 1997), it is unclear why similar patterns have not consistently been observed in shade-tolerant species of temperate forests (Canham et al. 1999, Lusk and Piper 2007). Interestingly, one aspect of our first hypothesis did receive support from one of our models: a negative impact of temperature on gap herbivory was found in tropical forests (Table 4). This result is consistent with insect avoidance of stress by radiative heating at high temperatures.

Meta-analysis supported our second hypothesis, that light environment influences insect herbivore pressure on plants through its effect on resource availability to herbivores (Richards and Coley 2008). However, although this effect was supported at a global scale, this result was entirely driven by strong patterns in tropical forests, the best-represented climate type in our compilation (Fig. 2, Table 1). Here, we found that the insect herbivory was significantly higher in sun than in shade. Higher herbivory in sun than in shade of tropical forests could reflect both greater plant growth and biomass in gaps (Bazzaz 1979), but also higher nutritional and energetic status of foliage. For instance, Richards and Coley (2008) found that the lepidopteran larva *Zunacetha annulata* feeding on sun leaves of its host plant *Hybanthus prunifolius*, ate 22 percent less leaf area, grew 25 percent faster, and had higher

pupal weights than larvae feeding on shade leaves. Although Richards and Coley (2008) found higher nitrogen concentration in shade leaves, both soil nitrogen mineralization and leaf nutrient status have been found to positively relate to light availability (Osier and Jennings 2007, Takafumi et al. 2010; but see Lusk and Reich 2000). The effect of gaps on plant growth and biomass could be more accentuated in the tropics, where understorey to gap-centre gradients of light availability are most pronounced (Ricklefs 1977, Lusk et al. 2011). In addition, faster leaf turnover in gaps than in the understorey could make gaps richer in availability of young leaves (Lowman 1992, Vincent 2006, Lusk and Corcuera 2011), which generally are more palatable than old leaves (Coley and Barone 1996). The strong effect of gaps on herbivory of shade-intolerant species (Fig. 3) is also consistent with our second (resource limitation) hypothesis. While shadeintolerant species exhibit significant biomass increments in response to increasing light availability, shade-tolerant species are less responsive (Bazzaz 1979, King 1994, Kitajima 1994). Thus, the nutritional benefit obtained by herbivores in gaps is expected to be much higher for shade-intolerant than for shade-tolerant species.

Higher herbivory in gaps could also occur if gaps attract fewer natural enemies of insect herbivores than the understorey. In a Puerto Rican montane tropical forest, the density of frogs that prey on a wide range of invertebrates was strongly reduced by experimental canopy opening (Klawinski et al. 2014). On the other hand, evidence from both tropical and temperate forests indicates that the impact of parasitism and predation on herbivores may be actually higher in gaps than in the neighbouring understorey (Harrison 1987, Richards and Coley 2007, 2008, Stoepler and Lill 2013). However, Richards and Coley (2007) reported that insect herbivory rates in tropical forests were higher in gaps than in understoreys, despite higher predator abundance in gaps. Spatial variation in pressure from natural enemies of insect herbivores therefore seems unlikely to explain higher rates of leaf damage in gaps than in understoreys.

Our findings have bearing on the long-standing question about latitudinal patterns of herbivory (MacArthur 1969, Coley and Aide 1991). In most studies examining

this question, herbivory has been evaluated under a similar light environment for the whole latitudinal range considered, attempting to standardize factors other than latitude. Our findings imply that patterns revealed by latitudinal comparisons of herbivory rates will depend on the light environment chosen by researchers. Regional differences in the effect of light environment on insect herbivory rates could account for the inconsistency among studies analysing latitudinal patterns of insect herbivory (Coley and Aide 1991, del-Val and Armesto 2010, Moles et al. 2011, Zhang et al. 2016).

Conclusions

Our study did not support the hypothesis that understorey insect herbivory in temperate forests (particularly in the Southern Hemisphere), is limited by low temperature. In contrast, our results are consistent with the hypothesis that the forest understorey limits resource availability for herbivores, especially in the tropics. The evidence that insect herbivory rates overall were higher in gaps than in understories suggests the selective importance of insect herbivory in forest understories may have been over-estimated. Our finding of latitudinal moderation of the effect of light environment on insect herbivory indicates that future investigations of latitudinal trends in herbivory should consider multiple light environments.

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Supplementary material (available online as Appendix oik-04686 at < www.oikosjournal.org/appendix/oik-04686 >). Appendix 1–2.

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